

Journal of Food and Dairy Sciences

Journal homepage & Available online at: www.jfds.journals.ekb.eg

Hepatotreatment Effect of Silybum, Spirulina and Propolis Consumption Compared to Silymarin Drug in Experimental Rats

Abd El-Ghany, M. A. ; Rasha M. Nagib and Hanan A. A Hashem*



Faculty of Specific Education, Home Economics Dept., Mansoura University –Egypt.



ABSTRACT

The purpose of this study was to investigate whether Silybum, Spirulina, and Propolis may have a therapeutic impact when compared to the silymarin medication. The results revealed that, Negative control showed significant lower in weight gain, feed efficiency ratio, serum HDL-c, total protein, albumin, globulin, catalase (CAT), superoxide dismutase (SOD) and Cytochrome P450 (CYP450) and significant higher in serum liver enzyme (ALT, AST, ALP, γ GT), total bilirubin, creatinine, urea, uric acid, cholesterol, triglycerides, LDL-c, VLDL-c and cholesterol /HDLc, LDLc/HDLc and malondialdehyde (MDA) compared to negative control. Administration of Silymarin, Silybum, Spirulina Propolis and mixture of them to liver injured rats for both the treated or preventive group showed a significant decrease in serum liver enzyme (ALT, AST, ALP, γ GT), total bilirubin, creatinine, urea, uric acid, cholesterol, triglycerides, LDL-c, VLDL-c and cholesterol /HDLc, LDLc/HDLc and (MDA) and significant increase in weight gain, feed efficiency ratio, HDL-c, total protein, albumin, globulin, CAT, SOD, and CYP450 in compared to positive control. Histopathological examination revealed that consumption of Silymarin, Silybum, Spirulina Propolis can lower pathological changes in injured. It can be recommended that consumption of Silymarin, Silybum, Spirulina Propolis and mixed them could manage treat liver injury rat by improve liver and renal function, lipids profiles, antioxidant enzymes and Cytochrome P450.

Keyword: Silymarin, Silybum, Spirulina Propolis Nutritional, liver, CCL4, rat.

INTRODUCTION

In the experimental study of hepatology, carbon tetrachloride CCl₄ is a hepatotoxic chemical due to trichloromethyl, which induces necrosis of hepatocytes, activates inflammation, and promotes the development of hepatic fibrosis (Santoyo *et al.*, 2006).

As shown by Jacobs *et al.*, (2002), Silymarin (milk thistle) could prevent the absorption of toxins into hepatocytes and regulate the action on cellular and mitochondrial membrane permeability (Anna *et al.*, 2020).

Spirulina, a unicellular filamentous algae belonging to the Oscillatoraceae family, is a cyanobacterium that includes a high amount of proteins as well as vital nutrients such as carotenoids, vitamins, and minerals and is mostly utilised as a food additive (Khan *et al.*, 2005b). It also contains numerous biological actions, including the prevention of anaemia, the reduction of medication genotoxicity, and the prevention of fatty liver disease, in addition to hypolipidemic and hypoglycemic effect. According to in vitro research, Spirulina has the potential to remove superoxide radicals and hydroxyl while also inhibiting lipid peroxidation (Mazo *et al.*, 2004 Premkumar *et al.*, 2004 and Li *et al.*, 2007).

Propolis is a resinous substance of diverse colours and textures that *Apis mellifera* bees collect from various plant sources. It mostly contains resins, flavonoids, phenolic aldehydes (polyphenols), waxes, fatty acids, essential oils, pollen, organic matter, and other minerals. It also contains vitamins B, C, and E, as well as essential minerals and trace elements (Hegazi *et al.*, 2000). Propolis thus possesses anti-inflammatory, antibacterial, antiviral, immunomodulatory,

antioxidant, and anti-proliferative activities (Almeida and Menezes, 2002).

Silybum marianum (*Cardus marianus*), commonly known as milk thistle or St. Mary thistle, is one of the oldest and thoroughly researched plants in the treatment of liver diseases (Pradhan and Girish, 2006). The active constituent of milk thistle is silymarin, a mixture of flavonolignans comprised of 4 isomers: silibinin, isosilibinin, silichristin, and silidianin. Most supplements are standardized according to their silibinin (often called silybin) content, the main component of the silymarin (Post-White *et al.*, 2007). Silymarin/silybin has been currently used for the treatment of cirrhosis, chronic hepatitis and liver diseases associated with alcohol consumption and environmental toxin exposure (Gazak *et al.*, 2004).

The purpose of this study Silybum, Spirulina, and Propolis could treat hepatic toxicity caused by carbon tetrachloride better than silymarin medication.

MATERIALS AND METHODS

Materials

1-Spirulina (biomass of cyanobacteria (blue-green algae) dried powder and silybum seeds (*Cardus marianus*) were obtained at a local herbal store in Cairo, and propolis powder was purchased from a unique beehive retail market in Beni Suef.

The Research Institutes of Ophthalmology in Giza, Egypt, provided 70 male white rats (181±8 g). Rats were placed in plastic mesh cages in an air-conditioned unit with a 12 hour light/dark cycle. Rats were fed a conventional food

* Corresponding author.

E-mail address: hananahmed113@yahoo.com

DOI: 10.21608/jfds.2022.120162.1037

and water ad libitum for a week before to experimentation to acclimate them. The experimental protocols were authorised in accordance with the Ethics Committee, Faculty of Specific Education, Mansoura University, Egypt's Guide for the Care and Use of Laboratory Animals.

2-According to NRC(1995), the usual diet consisted of maize starch (497g/kg), casein (200g/kg), sucrose (100g/kg), corn oil (50g/kg), cellulose (30 g/kg), mineral mixture (100g/kg), vitamins mixture (20g/kg), and DL-methionine (3g/kg) acquired from El-Gomhuria Company for Chemicals

3-Carbon Tetrachloride was purchased from El-Gomhuria Company for Chemicals in El-Mansoura, Egypt (3 ml/kg body weight, 50% in olive oil).

According to Lee *et al.*, the predicted dose for producing rat liver damage was 0.5ml/bw rat, delivered by back intraperitoneal injection (2005).

4-Silymarin (TMLegalon 140) tablets were acquired from El-Gomhuria Company for Chemicals in El-Mansoura City, Egypt, from a pharmacy.

Gamma Trade Company for Pharmaceutical and Chemicals, Dokki, Egypt, provided 5-kits.

Methods

Experimental protocol

Next the period of adaptation Rats were divided into seven groups at random. The first group only followed the standard diet (-V).

The other six groups were fed a standard diet and injected intraperitoneally with two doses of CCl4 before being reclassified as the control group (+ve), the treated groups fed 200 mg/kg bw silymarin, 400 mg/kg bw Silybum, 400 mg/kg body weight spirulina, 300 mg/kg body weight propolis, and 300 mg/kg body weight a mixture of the above agents (100 mg/kg silymarin, 200 mg and 150 mg/kg of Propolis) by gastric gavage every day for eight weeks. Chapman *et al.* calculated the daily food intake, weekly body weight gain, and feeding ratio (FER).

The serum enzymes activity of liver] alanine and aspartate amino transferase (ALT and AST), alkaline phosphatas (ALP) and gamma glutamyltransferase (GT), total bilirubin, total protein, albumin (A) creatinine, urea, and uric acid, and serum lipid profiles such as cholesterol, high-density lipoprotein cholesterol (HDLc) Serum superoxide dismutase (SOD), catalase (CAT), malondialdehyde (MDA), and cytochrome P450 levels were measured in accordance with Beuchamp and Fridovich. Sinha (1972), Draper and Hadley (1990), and Crespi *et al.*, (2002) are three examples. Serum globulin, low density lipoprotein cholesterol (LDLc), very low-density lipoprotein cholesterol (VLDLc), albumin/globulin ratio (A/G), and athrogenic indexes were calculated using Coles (1974), Lee and Nieman (1996), Friedewald *et al.* (1972), and Castelli and Levitar (1973).

Histological study:

The liver was extracted and fixed in 10% neutral buffered formalin, then cleaned in xylene and embedded in paraffin. 4-5um tick sections were produced and stained with Hematoxylin and Eosin (H&E) for future histopathological evaluation (Bancroft *et al.*, 1996).

Analyses statistical

SPSS software was used to conduct statistical analysis on the gathered data.

Calculations were made using analysis of variance ANOVA and subsequent LSD (SPSS). According to Artimage and Berry, computer programmes vary (1987).

RESULTS AND DISCUSSION

Carbon tetrachloride CCl4 has been shown to be hepatotoxic in rats.

Table (1) shows that the Control (+ve) group had lower final weight, weight growth (%), and FER (p0.001) than the Control (-ve) group. Overall, all treated rat groups exhibited substantial increases in final weight, weight growth, weight gain (percent) and FER (p0.001) in comparison to the control (+ve) group. There was no significant difference in final weight, weight growth, weight gain (percentage) and FER between the mixed group and the negative control group.

Table 1. Nutritional indications for hepatotoxic rat groups after treatment with diets enriched with silymarin, silybum, spirulina, propolis, and Their mixture.

Variables Groups	Final Weight(g)	Weight Gain(g)	Weight Gain%	Food Intake(g)	(FER)
Negative control group	a 228.50 ±7.47 d***	a 47.50 ±4.86 f***	a 26.34 ±2.81 f***	a 21.53 ±2.98 ab	a 0.036 ±0.013 d***
Positive control	193.80 ±3.61 c*	12.20 ±4.80 e***	7.32 ±2.39 e***	18.45 ±0.49 a	0.011 ±0.0065 cb***
Silymarin	206.50 ±4.11 ab	20.90 ±1.66 b**	11.33 ±1.02 b**	19.43 ±0.56 a	0.0111 ±0.008 bc**
Silybum	218.20 ±8.74 c*	36.00 ±1.76 d***	19.49 ±1.50 d***	19.97 ±0.25 a	0.021 ±0.0135 ab
Spirulina	205.80 ±5.34 bc*	24.30 ±1.70 c***	13.39 ±0.91 c***	19.25 ±0.54 a	0.030 ±0.0017 b**
Propolis	212.60 ±5.93 ab	30.00 ±2.36 a	16.43 ±1.28 a	19.49 ±1.50 a	0.025 ±0.008 a
Mixture	227.60 ±8.94 a	44.10 ±2.28 a	24.17 ±1.34 a	20.46 ±0.32 a	0.035 ±0.011 a

All data were expressed as mean ± SD. All data were expressed as mean ± SD. Significant with control group * p < 0.05 ** p < 0.01 *** p < 0.001. Mean values in each column having different superscript (a b,c,d,)were significantly different at P < 0.05 . Mixture: silymarin, silybum, spirulina, propolis

The effect of all studied extracts on food intake (FI), body weight gain ratio (BWG %) and feed efficiency ratio (FER) were illustrated in table (1) and figures 2 and 3. It could be observed that CCL4-positive control group rats group showed high significant decrease in body weight gain (g), body weight gain percentage (BWG %) and feed efficiency ratio (FER) compared to control negative group. El-Sayed et al (2012) Saber et al., (2011) and Aldo Ferreira et al., (2010). reported similar results this finding is due to hepatotoxic effect of CCL4 that affects weight gain in those rats but observed that all cirrhotic groups fed on basal diet with different levels of spirulina (0.25, 0.5 and 1%) had significant increase in body weight gain (BWG %) and food efficiency ratio (FER) compared with control positive rats. In agreement with Shaker et al., 2010 and Soufy, 2012 treatment with silymarin, silybum extract and propolis showed significant increase in final weight (g), weight gain (g), food intake (g) and FER, similar results were obtained on treatment with spirulina the same as results reported by Osman et al., 2012. Moreover, treatment with a mixture of all extracts showed significant

increase in final weight (g), weight gain (g), weight gain (%) food intake (g) and FER.

The data in table (2) showed that the control (+ve) group had a significant increase in serum AST, ALT, ALP, GT, and bilirubin values when compared to the control (-ve) group. However, when compared to the control (+ve), all treated groups silymarin, silybum, spirulina, propolis, and a combination of them showed a significant decrease in serum AST, ALT, ALP, GT, and bilirubin values. The results of the group fed a mixture of them showed a significant decrease in serum AST, ALT, ALP, GT, and bilirubin when compared to the treated groups.

Weber et al., (2003) investigated the CCl4 induced hepatic toxic accompanied by elevation in serum levels of AST, ALT,ALP, and GT enzymes with increased levels of serum bilirubin, creatinine, urea, and uric acid, lipids profile, and increase Oxidative stress, thereby confirming the typical hepato toxic effects of CCl4 in rats. These findings are consistent with those of Ozturk et al., (2012), Albassam et al., (2017), and Fenclova et al., (2019), who found that the ameliorating effect of silymarin and silybum on liver enzymes

(ALT, AST, and ALP) from hepatic damage induced by CCl4 was elevated in hepatotoxic rats, but rats treated with Silymarin or Silybum after CCl4 Saber et al., (2011) and Osman et al., (2012) discovered that spirulina appears to preserve the structural integrity of the hepatocyte cell membrane, as evidenced by a significant reduction in CCl4-induced elevation of serum enzymes. The decrease in serum enzymes could be attributed to their ability to prevent intracellular enzyme leakage via membrane stabi lising activity. Spirulina has been shown to have potent antioxidant and free radical scavenging properties. Spirulina's protective effect against CCl4-induced injury can be attributed to its high levels of antioxidants such as vitamins, carotenoids, and phycocyanin. In a similar study, Ozturk et al., (2012) agreed with our findings as they demonstrated significant elevations in serum creatinine, urea, and uric acid in rats administered only by CCL4 and reduced by consumption of silymarin. Demir et al., (2014) and You After treatment with diets supplemented with silymarin, silybum, spirulina, propolis, and a combination of these, the lipid profile of all hepatotoxic rats improved significantly.

Table 2. Serum liver enzymes and bilirubin levels in hepatotoxic male rats after treatment with diets enriched with silymarin, silybum, spirulina, propolis, and Their mixture.

Variables Groups	Serum liver enzymes and bilirubin levels							
	ALT m/ml	AST m/ml	ALP m/ml	γGT	Total Bilirubin	Direct Bilirubin	Indirect Bilirubin	
control -ve	f 42.99±2.59	f 31.63±3.51	a 155.07±12.05	f 25.0±4.69	c 0.61±0.12	d 0.46±0.03	e 0.15±0.04	
CCL4 induced-hepatotoxic groups	Positive control	a*** 125.46±12.09	a*** 193.84±13.87	b*** 88.50±8.6	a*** 270.08±14.29	a*** 3.86±0.28	a*** 1.79±0.16	a*** 2.07±0.16
	Silymarin	d** 72.85±7.62	d*** 122.41±1.61	d** 54.2±5.22	bc** 173.94±11.61	b** 1.44±0.11	bc* 0.79±0.09	c** 0.65±0.09
	Silybum	c*** 83.14±8.34	b*** 163.18±12.4	bc*** 64.9±6.79	b* 183.75±13.61	b** 1.38±0.17	bc** 0.75±0.11	c*** 0.63±0.16
	Spirulina	cd*** 74.18±7.43	bc*** 155.59±11.81	c*** 60.2±6.65	b* 181.32±11.94	b** 1.95±0.11	b** 1.02±0.13	b** 0.93±0.13
	Propolis	b*** 96.7±9.28	b*** 172.32±13.04	b*** 73.2±7.34	b** 192.15±14.18	b** 1.92±0.14	b** 1.06±0.16	bc** 0.86±0.17
	Mixture	de** 62.1±6.32	e** 74.97±5.5	de* 45.6±4.42	bc*** 172.6±16.61	c 0.85±0.11	cd 0.58±0.10	de 0.26±0.04

All data were expressed as mean ± SD. All data were expressed as mean ± SD. Significant with control group * p < 0.05 ** p < 0.01 *** p < 0.001. Mean values in each column having different superscript (a ,b, c,d,.) were significantly different at P < 0.05. Mixture: silymarin, silybum, spirulina, propolis

The data in Table (3) indicated that the control Positive rat group had a substantial rise in blood A/G ratio, creatinine, urea, and uric acid(p<0.001&0.05), but a significant reduction in serum total protein and globulins (p0.05). Additionally, all treated groups fed silymarin, silybum, spirulina, propolis, or a combination of them demonstrated a significant decrease in serum A/G ratio,

creatinine, urea, and uric acid, as well as a significant increase in serum total protein and globulin (p<0.001), when compared to the control positive group. The group given a combination of them had no significant difference in blood total protein, albumin, globulins, creatinine, urea, or uric acid levels as compared to the negative control group.

Table 3. Serum indicators of liver and kidney function in hepatotoxic rats fed diets enriched with silymarin, silybum, spirulina, propolis, and Their mixture.

Variables Group	Total Protein(g/dl)	Albumin (g/dl)	Globulin (g/dl)	(A/G) Ratio	Creatinine (mg/dl)	Urea (mg/dl)	Uric acid (mg/dl)	
Negative control group	a 7.15±0.64	a 4.01±0.32	a 3.14±0.34	c 1.21±0.29	e 0.46±0.03	b 35.03±3.22	b 2.47±0.85	
CCL4 induced- hepatotoxic groups	Positive control	b** 5.19±0.32	a 4.12±0.32	c*** 1.76±0.41	a*** 1.94±0.33	a*** 1.12±0.19	a*** 54.96±2.72	a*** 4.57±0.13
	Silymarin	a 7.06±0.60	a 4.09±0.38	b* 2.65±0.56	b* 1.63±0.52	d* 0.61±0.04	b 34.88±3.34	b 2.55±0.16
	Silybum	a 6.87±0.38	a 4.16±0.36	ab 2.7±0.63	b* 1.66±0.61	c** 0.72±0.05	b 33.58±3.01	b 2.50±0.08
	Spirulina	a 7.13±0.11	a 3.98±0.25	ab 2.8±0.44	b* 1.43±0.29	d* 0.63±0.3	b 33.98±3.46	b 2.50±0.12
	Propolis	a 6.88±0.31	a 4.12±0.42	ab 2.75±0.53	b* 1.57±0.46	b** 0.92±0.05	b 34.59±2.72	b 2.48±0.14

All data were expressed as mean ± SD. All data were expressed as mean ± SD. Significant with control group * p < 0.05 ** p < 0.01 *** p < 0.001. Mean values in each column having different superscript (a ,b, c,d,.) were significantly different at P < 0.05 Mixture: silymarin, silybum, spirulina, propolis

Similar findings were found in the study of EL-Sayed et al., (2012), who discovered that all cirrhotic

groups fed a basic diet containing varying levels of spirulina had significant reductions in total lipids, triglycerides, and

total cholesterol when compared to the positive control group of rats. Our findings appear to support the findings of Bashandy *et al.*, (2011), who discovered that spirulina significantly reduced HgCl₂-induced hepatotoxicity and modulated the lipid profile via its antioxidant properties. However, in a human study conducted by Aldo Ferreira *et al.*, (2010), it was discovered that spirulina extract significantly improved the serum lipid profile in man. Furthermore, Mahran *et al.*, 1996 and El Menyiy *et al.*, 2016 reported that serum total cholesterol, triglycerides, and LDLc were significantly increased while HDLc was significantly decreased in rats injured with thioacetamide (TAA) compared with control rats, the injured rats and fed on aqueous extracts of propolis could significantly modulate the lipid contents by decreasing total lipids profile levels with significantly increase in HDLc,

According to the data in table (4), the positive control group had a substantial rise in blood cholesterol, triglycerides, LDLc, VLDLc, and atherogenic index (p< 0.001), whereas the negative control group had a significant drop in serum HDLc (p< 0.001). While all treated groups fed silymarin, silybum, spirulina, propolis, or a combination of these showed significant decreases in serum cholesterol, triglycerides, LDLc, VLDLc, and atherogenic index (p 0.05 & p 0.01, respectively) and significant increases in serum

HDLc (p< 0.05) compared to the positive control group. In comparison to the negative control group, the group given a combination of them exhibited no significant difference in serum triglycerides, HDLc, or VLDLc.

Similar findings were found in the study of EL-Sayeda *et al.*, (2012), who discovered that all cirrhotic groups fed a basic diet containing varying levels of spirulina had significant reductions in total lipids, triglycerides, and total cholesterol when compared to the positive control group of rats. Our findings appear to support the findings of Bashandy *et al.*, (2011), who discovered that spirulina significantly reduced HgCl₂-induced hepatotoxicity and modulated the lipid profile via its antioxidant properties. However, in a human study conducted by Aldo Ferreira *et al.*, (2010), it was discovered that spirulina extract significantly improved the serum lipid profile in man. Furthermore, Mahran *et al.*, 1996 and El Menyiy *et al.*, 2016 reported that serum total cholesterol, triglycerides, and LDLc were significantly increased while HDLc was significantly decreased in rats injured with thioacetamide (TAA) compared with control rats, the injured rats and fed on aqueous extracts of propolis could significantly modulate the lipid contents by decreasing total lipids profile levels with significantly increase in HDLc,

Table 4. Serum lipids profile of hepatotoxic rats fed diets enriched with silymarin, silybum, spirulina, propolis, and Their mixture.

Variables Groups	Cholesterol (mg/dl)	Triglycerides (mg/dl)	LDLc (mg/dl)	HDLc (mg/dl)	VLDLc (mg/dl)	Cholesterol /HDLc	LDLc/ HDLc
Negative control	d 58.56±8.59	cd 30.81±5.75	e 20.17±1.33	a 32.23±2.52	cd 6.16±0.75	d 1.82 ± 0.25	D 0.63 ± 0.25
Positive control	a*** 112.68±6.94	a*** 111.70±6.43	a*** 79.1±7.86	d*** 11.24±1.87	a*** 22.34±1.28	a*** 10.02 ± 1.77	a*** 7.03 ± 1.45
Silymarin	bc* 85.12±3.06	c 39.24±5.56	b*** 54.58±4.37	bc* 22.69±1.38	cd 7.84±0.51	bc** 3.79 ± 0.34	b** 2.40 ± 0.33
Silybum	bc* 86.95±2.39	c 40.26±5.49	bc*** 51.71±7.78	ab 27.19±3.11	c 8.05±0.49	c** 3.19 ± 0.33	bc** 1.90 ± 0.31
Spirulina	bc* 84.57±2.28	c 40.17±6.28	b*** 53.58±5.30	b* 22.97±1.32	c 8.03±0.45	c** 3.69 ± 0.33	b** 2.33 ± 0.16
Propolis	b** 94.41±2.78	b** 66.40±8.90	b*** 57.97±3.32	b* 23.16±3.72	b* 13.28±1.78	b** 4.01 ± 0.50	b** 2.50 ± 0.31
Mixture treated	bc* 84.03±1.43	c 38.94±5.0	cd** 49.15±1.02	ab 27.09±3.81	c 7.78±0.40	c** 3.1 ± 0.66	bc** 1.81 ± 0.68

All data were expressed as mean ± SD. All data were expressed as mean ± SD. Significant with control group * p < 0.05 ** p < 0.01 *** p < 0.001. Mean values in each column having different superscript (a, b, c, d, e) were significantly different at P < 0.05 Mixture: silymarin, silybum, spirulina, propolis

The data in table (5) indicated that the control positive rat group had a substantial rise in serum malondialdehyde (MDA) levels but a significant reduction in serum catalase, superoxide dismutase (SOD), and cytochrome CYP450 levels (p0.001). On the other hand, all treatment groups given silymarin, silybum, spirulina, propolis, or a combination of them had a substantial rise in serum catalase, SOD, and CYP450 levels, but a significant drop in serum MDA levels (p0.01), as compared to the control positive group.

It is now generally accepted that oxidative stress due to increased ROS production has a role in the pathogenesis of hepatic injury induced by CCL4. Hepatocytes are continuously exposed to ROS and are protected from oxidative injury by a range of antioxidant pathways (Zhang and Morris., 2003). The state of oxidative stress exists when there is an imbalance between pro-oxidant and antioxidant chemical species.

In the present study, it was observed that there was a significant decrease in SOD and catalase activities with

significant increase in MDA (marker of lipid peroxidations) in the serum of rats administered with CCL4 only. Moreover, treatment of rats with spirulina extracts, silymarin, silbum marianum and propolis caused significant improvement in the serum levels of catalase and SOD activities and MDA compared to positive control group. Qureshi *et al.*, 2007 demonstrated significant reduction in liver content of GSH with significant elevation of lipid peroxidation in CCL4-treated rats.

In agreement with our study Osman *et al.*, 2012 proved the antioxidant effect of spirulina against hepatotoxicity, that was due to the anti-inflammatory and anti-oxidative properties of its constituent phycocyanin a biliprotein found in spirulina that exerts a scavenging action against reactive oxygen species as well as anti-inflammatory activity, also spirulina has other antioxidant and anti-inflammatory agents such as β carotene, phenolic compounds.

Table 5. Serum levels of many antioxidant enzymes and Cytochrome P450 (CYP450) in the negative control and hepatotoxic rat groups after treatment with diets supplemented with silymarin, silybum, spirulina, propolis, and a Their mixture.

Serum levels of many antioxidant enzymes and Cytochrome P450 (CYP450)					
Variables Group	MDA (Nmol/l)	CAT (U/ml)	SOD (U/ML)	CYP450	
Negative control	c 2.60 ±0.71	a 28.00 ±2.58	a 1047.20 ±51.25	a 20.62 ± 1.48	
Positive control	a*** 13.48 ± 1.13	e*** 3.90 ±0.48	fe*** 18.45 ± 0.49	e*** 1.11 ± 0.24	
CCL4 induced-hepatotoxic groups	Silymarin	b* 4.95 ±0.34	c*** 18.47 ±1.06	b** 846.39 ±24.95	a 19.05 ± 1.05
	Silybum	b* 5.32 ±0.48	cd** 15.78 ±2.01	c* 741.31 ±23.35	bc* 14.2 ± 0.48
	Spirulina	b* 5.30 ±0.45	c** 181.1 ±1.33	b** 837.97 ±30.07	b** 15.77 ± 0.71
	Propolis	b* 5.73 ±0.59	c** 16.35 ±2.40	d** 637.45 ±26.75	d** 12.13 ± 0.58
	Mixture	c 2.57 ±0.73	b* 22.43 ±1.03	e *** 473.45 ±34.12	cd** 13.4 ±1.21

All data were expressed as mean ± SD. All data were expressed as mean ± SD. Significant with control group * p < 0. 05 ** p < 0. 01 *** p < 0.001 Mean values in each column having different superscript (a ,b, c,d,) were significantly different at P < 0.05 Mixture: silymarin, silybum, spirulina, propolis

Concerning the antioxidant effect of aqueous propolis extract El-Khatib *et al.*, 2002 published results similar to our results in the current study where they found

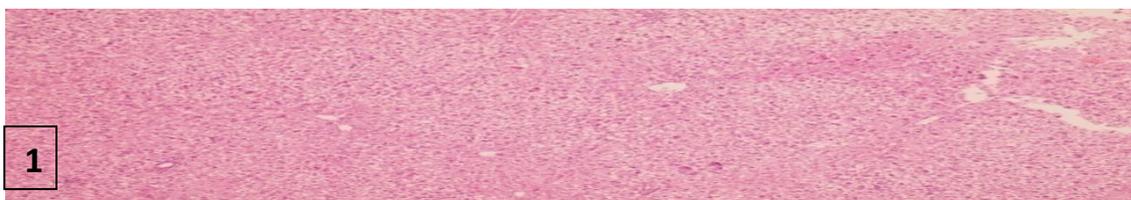
that treatment with aqueous propolis extract normalize lipid peroxidation.

Shaarawy *et al.*, 2009 induced hepatic destruction by N-nitrosodiethylamine (NDEA) plus CCl4. In their experiment, liver function enzymes and hepatic lipid peroxidation (LPO) was found to be increased and superoxide dismutase (SOD), and GSH-dependent enzymes decreased. Pradhan and Girish 2006 also showed that another N-nitroso ALPyI compound, Diethylnitrosamine, induced alterations in the liver tissue and silymarin reversed these negative changes by improving antioxidant capacity. Protective effect of silymarin has also been shown on hepatic damage by many agents such as ethanol (Loguercio and Federico, 2003) and anti-tuberculosis drugs (Gazak *et al.*, 2004).

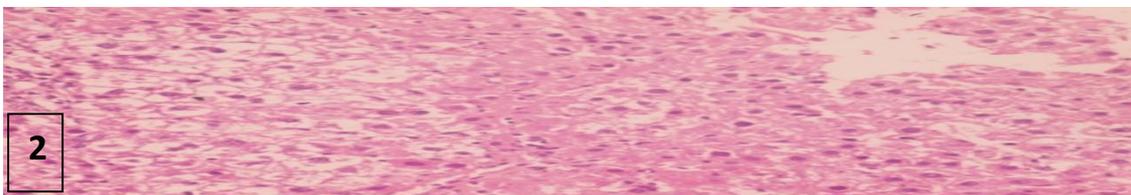
Varga *et al.*, 2004 made hepatotoxicity by cisplatin (CDDP) administration, which is a widely used anticancer drug. They showed that silymarin attenuated serum ALT, AST, liver nitric oxide (NO) and malondialdehyde (MDA) level, previously increased by CDDP.

Liver Examination Histopathological Findings

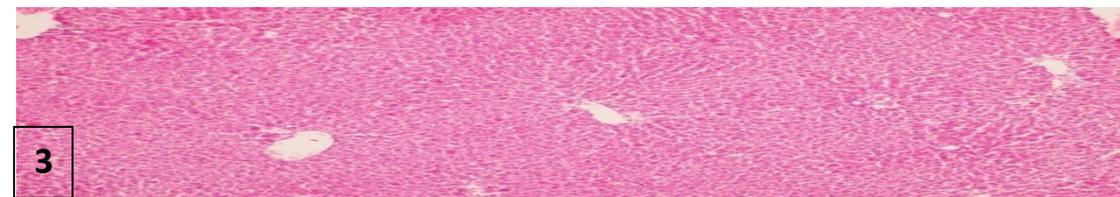
The negative group's liver tissue exhibited the usual histological structure of the hepatic lobule (pic 1). Intact liver architecture—intact central vein or portal tract—no steatosis observed—no necroinflammatory damage identified Microscopically, the liver tissue of the positive control (+ve) demonstrated a region of coagulative necrosis in hepatocytes with infiltration of mononuclear inflammatory cells, cytoplasmic vacuolization of hypatocytes, necrosis of sporadic hepatocytes, and inflammatory damage (pic 2) Examined slices of liver tissue from all injured rats treated with silymarin, silybum, spirulina, propolis, or a combination of these herbs revealed a virtually normal histological structure in pictorial form (3, 4, 5, 6, 7)



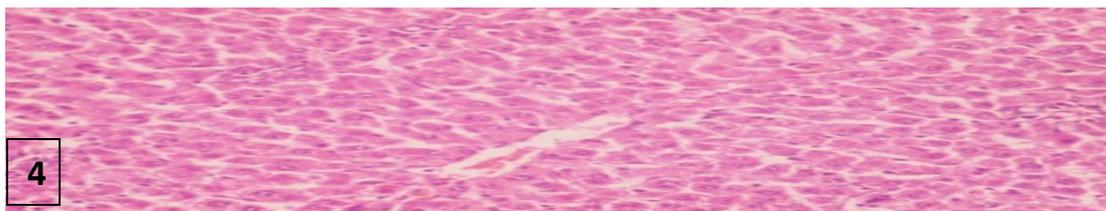
Pict 1. liver specimens showing a=loss of normal liver architecture vacuolated hepatocytes, focal necrosis and sinusoidal congestion (CCL4-untreated rats group, x100).



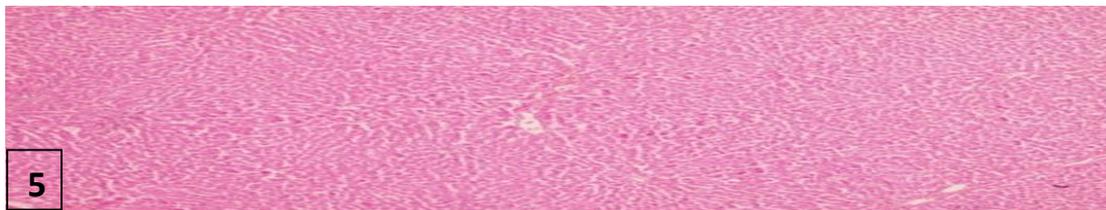
Pict 2. area of coagulative necrosis in hepatocytes with mononuclear inflammatory cells infiltration (positive control (CCL4-untreated rats group, x400),



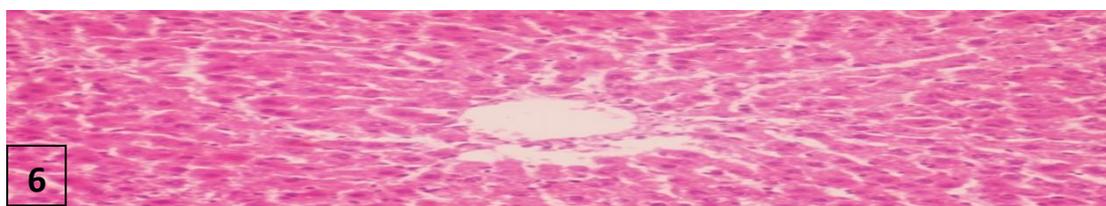
pict 3. nearly normal liver architecture with normal hepatocytes and Kupffer cells (Silymarin-treated group, x100).



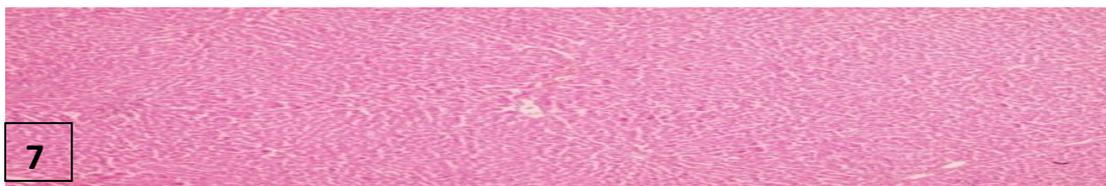
Pict 4. Liver specimens showing a= normal central vein, normal hepatocytes, hepatic cords, sinusoids and Kupffer cells (*Silybum*-treated group, x400).



Pict 5. normal liver architecture with mild sinusoidal congestion (*Spirulina*-treated group, x100).



Pict 6. Liver specimens showing a= portal regeneration of hepatocytes with residual degeneration with marked infiltration of sinusoids with mononuclear inflammatory cells (*Propolis*-treated group, x400).



Pict 7. Totally normal liver architecture with regenerated hepatocytes and normal Kupffer cells (mixture-treated group, x400)

The microscopic examination of liver from CCL4 administered untreated rats revealed a more severe and persistent change in degeneration, swelling of hepatocytes, loss of liver tissue structure, severe dilatation and vascularization, indicating liver injury (Picture 2). This result was previously published by El-Khatib *et al.*, (2002), Shaker *et al.*, (2010), and Soufy, (2012). Furthermore, treatment with silymarin, silybum (pict.3-4-5-6). In agreement with these pathological findings, Ozturk *et al.*, (2012) and Abenavoli *et al.*, (2018) discovered that treatment with silbum marianum and silymarin significantly reduced the liver histopathological findings. EL-Sayeda *et al.*, (2012) and Hotta *et al.*, (2020) discovered that microscopic examination of the livers of fibrotic rats fed a basic diet containing spirulina (0.25 and 0.5 percent) with subcutaneous injection of CCl4 shows mild degenerative change and mild swelling of hepatocytes. Ours is the first study to show a significant improvement in serum liver and renal function, lipid profile, antioxidant enzyme, and CYP450 by a combination of spirulina, album, propolis, and silymarin.

REFERENCES

Abenavoli, L.; Izzo, A.A.; Milić, N.; Cicala, C.; Santini, A. and Capasso, R. (2018): Milk thistle (*Silybum marianum*): A concise overview on its chemistry, pharmacological, and nutraceutical uses in liver diseases. *Phyto- Res*, 32 (11), 2202-2213.

Albassam, A. A.; Frye, R.F. and Markowitz, J.S. (2017): The effect of milk thistle (*Silybum marianum*) and its main flavonolignans on CYP2C8 enzyme activity in human liver microsomes. *Chem-biological inter*, (271): 24-29.

Aldo Ferreira-Hermosillo, Patricia V Torres-Duran and Marco A Juarez-Oropeza (2010): Hepatoprotective effects of *Spirulina maxima* in patients with nonalcoholic fatty liver disease: a case series. *Journal of Medical Case Reports*, 4:103.

Almeida, E.C. and H. Menezes, (2002): Anti-inflammatory activity of propolis extracts: a review. *J. Venom. Anim. Toxins.*, 8: 191-212.

Anna, E., Krisztina, S.S., Anna, B.E., Fehérd, K.H and Hedvig, F. (2020): Impact of milk thistle (*Silybum marianum*) on the mycotoxin caused redox-homeostasis imbalance of ducks live. *J* (187): 181-187.

Artimage, G.Y. and Berry, W.G. (1987): *Statistical Methods* 7th Ed. Ames, Iowa State University Press, 39-63.

Bancroft, J.D, Stevens, A. and Turner, D. R. (1996): *Theory and practice of histological technique* .4th Ed, New York, Churchill, Livingstone.

Bartholomev, R.J. and Delany, A. (1966): *Proc Aust. Assoc .Biochemists* 1, 214.

- Bashandy S. A.1., Alhazza I. M., El-Desoky G. and Al-Othman Z. (2011): Hepatoprotective and hypolipidemic effects of *Spirulina platensis* in rats administered mercuric chloride. *African Journal of Pharmacy and Pharmacology*. 5(2), 175-182.
- Beuchamp, C. and Fridovich, J. (1971): Superoxide dismutase. Improved assay and assay applicable to acrylamide gels. *Anal Biochem.*, 44: 276-287.
- Bonsens, K. E. and Taussky, D. H. (1984): Determination of serum creatinine. *J Ch Inv*, 27: 648-660
- Buccolo, G. and David, H. (1973): Quantitative determination of serum triglycerides by use of enzymes. *Clin. Chem.*, 19: 419-32.
- Castelli, T. and Levitar, Y. (1977): Atherogenic index. *Curr Presc* p39.
- Chapman, D.G., Gastilla R. and Campbell, T.A. (1950): Evaluation of protein in food. I. A. Method for the determination of protein efficiency ratio. *Can. J. Biochem. Physio.*, 1 (37) 679-686.
- Coles, E. H. (1974): *Veterinary Clinical Pathology*. Saunders Company, Philadelphia and London.
- Creşpi C.L., Miller V.P. and Stresser D.M. (2002): Design and Application of Fluorogenic Assays for Human Cytochrome P450 Inhibition. In: Johnson E. F., Waterman M. R., editors. *Cytochrome P450, Part C. Vol 357*. London: Academic Press;
- Draper, H. H. and Hadley, M. (1990): Malondialdehyde determination as index of lipid peroxidation, *Methods Enzymol.* 186, 421-431.
- El Menyiy N., Al Waili N., Bakour M., Al-Waili H., and Lyoussi B. (2016): Protective effect of propolis in proteinuria, crystaluria, nephrotoxicity and hepatotoxicity induced by ethylene glycol ingestion. *Arch Med Res* 47(7):526-534.
- El-Khatib S., Agha M., Mahran G. and Khayyal T. (2002). Prophylactic effect of Aqueous Propolis extract against acute experimental hepatotoxicity in vivo. *Naturforsch.* 57: 379-385.
- EL-Sayeda, G. E. EL-Sahar and Abor, M. M. Abed EL-Rahman. (2012): Hepatoprotective Activity of Different Doses of *Spirulina* against Ccl4 Induced Liver Damage in Rats] *J Am Sci*;8(8):916-923.
- Fenclova, M.; Novakova, A.; Viktorova, J.; Jonatova, P.; Dzuman, Z.; Ruml, T.; Kren, V.; Hajslova, J.; Vitek, L.; and Stranska-Zachariasova, M. (2019): Poor Chemical and Microbiological Quality of the Commercial Milk Thistle-Based Dietary Supplements May Account for Their Reported Unsatisfactory and Non-Reproducible Clinical Outcomes. *Sci. Rep.*, 9, 11118.
- Fossati, P.; Prencipe, L. and Berti, G. (1980): Use of 3, 5-dichloro-2-hydroxybenzene sulfonic acid /4-aminophenazon chromogenic system in direct enzymatic assay of uric acid in serum and urine. *Clin. Chem.*, 26: 227-231.
- Friedewald, W.T.; Levy, R.I. and Fredrickson, D.S. (1972): Estimation of concentration of low-density lipoproteins separated by three different methods. *Clin. Chem.* 28: 2077
- Grodon, T. and Amer, M. (1977): Determination of HDL. *J. Med.*, 62: 707.
- Hegazi, A.G. (2000): Propolis: An overview. *Congreso Internacional de propolis*. Durante los dias 1 y 2 de Septiembre de 2000 en Buenos Aires - Argentina
- Hotta S.; Uchiyama S. and Ichihara, K. (2020): Brazilian red propolis extract enhances expression of antioxidant enzyme genes in vitro and in vivo. *Biosci Biotechnol Biochem* 84(9):1820-1830
- Khan, Z.; Bhadouria, P. and Bisen P.S. (2005b): Nutritional and therapeutic potential of *Spirulina*. *Curr Pharm Biotechnol*, 6:373-379
- Kind, P.R. and King, E.J. (1954): Estimation of ALP alkaline phosphatase activity by determination of hydrolyzed phenol with aminoantipyrine. *J. Clin. Pathol.* 7, 322
- Lee, G.P.; Jeong, W.I.; Jeong, D.H.; Do, S.H.; Kim, T.H. and Jeong, K.S. (2005): Diagnostic evaluation of carbon tetrachloride-induced rat hepatic cirrhosis model. *Anticancer Res.*, Mar-Apr; 25(2A):1029-38.
- Lee, R. and Nieman, D. (1996): *Nutritional Assessment*. 2nd Ed. Mosby, Missouri, USA.
- Li, A.H.; Cheng, K.; Wong, C.; King-Wai, F.; Feng, C. and Yue, J., (2007): Evaluation of antioxidant capacity and total phenolic content of different fractions of selected microalgae. *Food Chemistry*, 102: 771-776.
- Mahran, L. G.; El-Khatib, A. S.; Agha A. M. and Khayyal. (1996), The protective effect of aqueous propolis extract on isolated rat hepatocytes against carbon tetrachloride toxicity. *Drugs Exp. Clin. Res.* 22, 309-316.
- Mazo, V.K.; Gmoshinskii, I.V. and Zilova I.S. (2004): Microalgae *Spirulina* in human nutrition. *Vopr. Pitan.*, 73:45-53
- Neuschwander-Tetri BA, Unalp A, Creer MH, *et al* (2008): Influence of local reference populations on upper limits of normal for serum alanine aminotransferase levels. *Arch Intern Med.*;168(6):663-6.
- NRC (1995): National Research Council: Nutrient requirements of laboratory animals. fourth revised edition, PP.29-30 National Academy Press. Washington, DC.
- Osman, H.; El-Sheekh, M.; Farrag, H.; El-Saeed M. and Kassem A. (2012): Hepatoprotective effect induced by NaCl-stressed *Spirulina platensis*: Histopathological, biochemical and histochemical studies. *World Applied Science Journal* 18 (10): 1370-1380.
- Ozturk, M.; Akdogan, M.; Keskin, I.; Kisioglu, A. N.; Oztas, S. and Yildiz, K. (2012): Effect of *Silybum marianum* on acute hepatic damage caused by carbon tetrachloride in rats. *Biomedical Research (0970-938X)*. Apr-Jun, Vol. 23 Issue 2, p268-274. 7p. 1
- Patton, C.J. and Crouch, S.R. (1977): Enzymatic calorimetric method to determine urea in serum. *Anal Chem.*, 49: 464-469.
- Premkumar, K.; Abraham, S.K.; Santhiya, S.T. and Ramesh A. (2004): Protective effect of *Spirulina fusiformis* on chemical-induced genotoxicity in mice. *Fitoterapia*, 75:24-31.
- Reitman, S. and Frankel, S. (1957) A calorimetric method for the determination of serum GOT and GPT. *American Journal of Clinical Pathology* 28:56-63.

- Reitman, S. and Frankel, S. (1957): Determination of glutamate pyruvate transaminase and glutamate oxaloacetate transaminase. Amer. J. Clin. Path., 28:56-63
- Richmond, W. (1973): Enzymatic determination of cholesterol. Clin. chem., 19: 1350-1356
- Saber A. Sakr, Sabah F. El-Abd, Mohamed Osman, Asmaa M. Kandil and Mona S. Helmy (2011): Ameliorative Effect of Aqueous Leave Extract of Ocimum Basilicum on Ccl4 – Induced Hepatotoxicity and Apoptosis in Albino Rats. Journal of American Science; 7 (8)
- Santoyo, S.; Herrero, M.; Javier, F.; Cifuentes, A.; Ibanez E. and Jaime L. (2006): Functional characterization of pressurized liquid extracts of Spirulina platensis. European. Food Research Technology, 224: 75–81.
- Shaker, E.; Mahmoud, H. and Mnaa S. (2010): Silymarin, the antioxidant component and *Silybum marianum* extracts prevent liver damage. Food and Chemical Toxicology 48, 803–806.
- Sinha, A.K. (1972): Colorimetric assay of catalase. Anal. Biochem., 47: 389–394
- Soufy, N. (2012). Hepatoprotective and Antioxidant Effects of Silybum Marianum Plant against Hepatotoxicity Induced by Carbon Tetrachloride in Rats. Journal of American Science; 8(4), 479-486
- Weber, L.W.; Boll, M.; Stampfl, A. (2003): Hepatotoxicity and mechanism of action of haloalkanes: Carbon tetrachloride as a toxicological model. Crit Rev Toxicol 33:105–136.
- Weichselbaum, T.F. (1946): An accurate and rapid method for the determination of protein in small amount of blood serum and plasma. Am J Clin Path (16):40.
- Yousif, A. El Hassanen I, Hanaa, B. Abeer N.A Abd EL-Rahman, and Naglaa M. B (2021): Potential Effect Of Milk Thistle (*Silybum Marianum*) On Liver Disorders Induced By Carbon Tetrachloride J of Home Economics(31):83-93

التأثير العلاجي للسمية الكبدية لتناول السيليبيوم والسبيرولينا والبروبوليس بالمقارنة بعقار السيليمارين في فئران التجارب عبد الغني محمود عبد الغني ، رشا محمد نجيب و حنان احمد احمد هاشم قسم الاقتصاد المنزلي بكلية التربية النوعية جامعة المنصورة

تهدف الدراسة لمعرفة التأثير العلاجي والوقائي للسيليمارين، والسيليبيوم والسبيرولينا والبروبوليس ومخلوطهم على الإصابة الكبدية في الفئران أظهرت نتائج المجموعة الضابطة الموجبة انخفاض معنوي في قيم الزيادة في الوزن، ونسبة كفاءة التغذية ومستوي السيرم من عالية الكثافة والبروتين الكلي والألبومين والجلوبيولين وسوبر أكسيد ديسموتاز والكتالاز والسيتوكروم وزيادة معنوية في وظائف الكبد والبيلروبين والكرياتينين واليوريا وحمض اليوريك والكوليسترول والدهون الثلاثية والليبيروتينات منخفضة الكثافة ومنخفضة الكثافة جدا ومؤشر تصلب الشرايين والمانولدهيد بالمقارنة بالمجموعة الضابطة السالبة كما أظهرت النتائج عند تناول كل من سيليمارين و سيليبيوم و سبيرولينا والبروبوليس وخلطهم للفئران المصابة بالكبد للمجموعة المعالجة والوقائية انخفاض معنوي في إنزيم الكبد والبيلروبين ووظائف الكلى الكرياتينين واليوريا وحمض اليوريك والكوليسترول والدهون الثلاثية والليبيروتينات منخفضة الكثافة ومنخفضة الكثافة جدا ومؤشر تصلب الشرايين والمانولدهيد وزيادة معنوية في زيادة الوزن ونسبة كفاءة التغذية والليبيروتينات مرتفعة الكثافة والبروتين الكلي والألبومين والجلوبيولين والكتالاز والسوبر أكسيد ديسموتاز والسيتوكروم بالمقارنة بالمجموعة الضابطة الموجبة كما أظهرت نتائج الفحص الهستولوجيا أن تناول كل من سيليمارين و سيليبيوم و سبيرولينا والبروبوليس ومخلطهم خفض من التغيرات المرضية للانسجة وتوصي الدراسة بضرورة تناول السيليمارين وسيليبيوم وسبيرولينا والبروبوليس ومخلطهم لعلاج الإصابة الكبدية لتحسين وظائف الكبد والكلى ودهون الدم ومضادات الأكسدة. و السيتوكروم

الكلمات المفتاحية: السيليبيوم-السبيرولينا-البروبوليس- السيليمارين-رابع كلوريد الكربون .