

Comparative Study on some Plants as Sources of Natural Antioxidants

Eman S. El-Ashaal¹; A. A. El-Refai² and Amal M. El-Bastawesy¹

¹Food Technology Research Institute, Agric. Res. Center, Giza, Egypt.

²Food Industries Dept., Fac. of Agric., Mansoura Univ., Egypt.



ABSTRACT

Nowadays, plants and natural products have a great probable interest due to their pharmaceutical, cosmetic and nutritional applications. So, the biologically active compounds of moringa and fig leaves as well as red pepper fruits and seeds were assessed as antioxidants agents as well as their antioxidant activities were evaluated using different assays and the obtained results revealed that, moringa and fig leaves had the highest content of chlorophylls A and B, but their contents in moringa leaves (625.37 and 229.94 mg/100g, respectively) were more than four folds greater than fig leaves (142.14 and 83.57 mg/100g, respectively). Red pepper fruits and moringa leaves had the highest contents of total carotenoids. The ABTS and DPPH procedures gave the highest antioxidant capacity values of all selected raw materials. Furthermore, moringa and fig leaves recorded the highest contents of antioxidant activity comparing with red pepper fruits and seeds correlated with the highly contents of total phenols and flavonoids. Moringa leaves had higher contents of phenolic acids (309.4mg/100g) than other selected plants and E- vanillic and pyrogallol were the most common phenolic compounds found in moringa leaves. On the other hand, there are little differences between fig leaves and red pepper fruits but, the lowest phenolic acids content (136.58 mg/100g) was detected in red pepper seeds. There was variation in flavonoids individuals between all selected raw materials. Moringa and fig leaves had approximately the same highest contents of flavonoids (283.71 and 266.73 mg/100g, respectively) than red pepper fruits and seeds which recorded lowest ones (112.84 mg/100g). Cobalamin (B₁₂) was the major B complex vitamins found in fig leaves, red pepper fruits, moringa leaves and red pepper seeds which recorded 994.01, 217.46, 164.81 and 125.5 mg/100g, respectively. Moringa leaves and red pepper fruits are considered as excellent sources for vitamins C and K that play an important roles as antioxidant agents to protect biological systems via inhibition or prevention of oxidation stress induced by reactive oxygen substances generated from normal metabolic activity or environmental factors followed by red pepper seeds and fig leaves.

Keywords: Moringa leaves, Fig leaves, Red pepper fruits and seeds, antioxidants activity assays

INTRODUCTION

Medicinal plants and natural products have been recognized as a source for medicines since ancient times across the world for treating and preventing human diseases. Many antioxidant compounds from plants such as carotenoids, ascorbic acid, flavonoid and phenolic compounds are the effective nutrients to prevent oxidative stress related disease including diabetes, Alzheimer's, Parkinson's, cardiovascular diseases, cancers and aging process (Volko *et al.*, 2007). Among the diversity of antioxidant constituents from plants, phenolic and flavonoid compounds have received a great deal of attention as antioxidant to prevent diseases caused as a result of oxidative stress (Karimi *et al.*, 2011).

Moringa (*Moringa oleifera* Lam), is a high source for in nutrients and phytochemical compounds as vitamins, proteins, β -carotene, amino acids and various phenolics such as caffeoylquinic acid, kaempferol, quercetin and zeatin with potential for nutritional and therapeutic applications (Siddhuraju and Becker, 2003). Different parts of this plant such as roots, leaves, bark, flowers, fruit of immature pods and seeds have a number of therapeutic properties such as, antioxidant, anti-inflammatory (Mahajan *et al.*, 2009), antihypertensive, antitumor, antibacterial (Rahman and Sheikh, 2009), cholesterol lowering, hepatoprotective (Mehta *et al.*, 2003) and are being operational in various traditional medicine system for curing different health problems (Stohs and Hartman, 2015).

Fig (*Ficus carica*) is a rich fruit containing vitamins, minerals, fats and having the highest plant sources for calcium, potassium (that help control blood pressure) and fibers. They are good sources of flavonoids, polyphenols and some bioactive compounds (Gilani *et al.*, 2008). The leaves of this plant have anti diabetic properties and reduce the amount of insulin needed by diabetics (Perez *et al.*, 2003), inhibit the growth of cancer cells and prevent colon cancer (Herre *et al.*, 2008).

Peppers (*Capsicum annum*) are a good source for several health-promoting compounds such as flavonoids, carotenoids, vitamin C and capsaicinoids, responsible for their characteristic hot taste, (Kim *et al.*, 2010). In peppers, there are phytochemical properties that have many biochemical and pharmacological properties which includes antioxidants, anti-inflammatory and anti-carcinogenic activities (Nishino *et al.*, 2009), antimicrobial activity (Wahba *et al.*, 2010) that can also boost immune system. Accordingly, this investigation was planned to evaluate the phytochemicals and bioactive compounds of selected raw materials as antioxidants agents as well as evaluate their antioxidant activities using different assays.

MATERIALS AND METHODS

Materials

Raw Materials

- Moringa seeds (*Moringa oleifera* Lam) were sown in experimental station of Agriculture of Faculty, Mansoura University during seasons of 2015 and 2016, then moringa terminal buds were used. Fig buds (*ficus carica*) were obtained from El Minia city and red hot pepper fruits (*Capsicum annum* L.) were obtained from a culture near Giza governorate, Egypt.
- Most chemicals (analytical grade) were purchased from Elgomhouria pharmaceuticals Co., Cairo, Egypt.
- 2,2-diphenyl-1-picryl-hydrazyl (DPPH), Folin-Ciocalteu reagent, 2,2-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid (ABTS), gallic acid, quercetin and butelated hydroxy toluene (BHT) were obtained from Sigma-Aldrich Chime, Steinheim, Germany.

Methods

Collection and preparation of plant materials

The fresh leaves of moringa and fig buds were collected washed in running tap water to remove impurities, drained from excess water and shade dried for 4 days. The dried leaves were grounded to fine powder (Anjorin *et al.*, 2010). The red pepper fruits were washed several times with tap water, the seeds and peduncle were removed, cut into small pieces, steamed for 5 min. and

blotted with tissue paper to remove excess water, dried in oven at 50±1°C for two days and ground to fine powder (Tunde-Akintunde, 2010). Whereas, the seeds and peduncles were removed, then dried alone in oven at 50±1°C for two days and ground to fine powder.

Analytical methods

- Total anthocyanins were determined according to the method described by Ranganna (1977). Both chlorophylls (A and B) and total carotenoids of samples were determined according to the methods described by Holm (1954) and Wetstein (1957).
- Total phenolic compounds content was determined using Folin-Ciocalteu reagent according to the method described by Maurya and Singh (2010) and phenolic compounds were fractionated and identified by HPLC according to the method described by Goupy *et al.* (1999).
- Total flavonoids content was determined according to the method described by Jia *et al.* (1999) and flavonoid compounds were fractionated and identified by HPLC according to the method of Loon *et al.* (2005).

Determination of antioxidant activity using different procedures

- The 2, 2'-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid (ABTS) assay was carried out according to Re *et al.* (1999).
- The antioxidant activity of samples was determined by the 2, 2'-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity according to the colorimetric method of Brand-Williams *et al.* (1995), as well as it was also determined by measuring the inhibition of hydroperoxides formed from linoleic acid oxidation (Dapkevicius *et al.*, 1998). The ferric reducing property of sample extracts was determined according to the method described by Lim and Murtijaya (2007).

- B- Complex vitamins were fractionated according to the method described by Batifoulier *et al.* (2005).
- Vitamin C was determined using 2, 6 Dichlorophenol - Indophenol according to the method described by Ranganna (1977).
- Vitamins A, D, E and K were determined according to the methods of Pyka and Sliwiok (2001).

The statistical analysis

The statistical analysis was carried out using one way analysis of variance (ANOVA) under significant level of 0.05 for the whole results using the statistical program CoStat (Ver. 6.400) and data were treated as complete randomization design according to Steel *et al.* (1997). To ascertain the significant among means of different samples, LSD test was applied.

RESULTS AND DISCUSSION

Bioactive compounds content of selected raw materials

The biologically active compounds of moringa and fig leaves as well as red pepper fruits and seeds were evaluated and the results are presented in Table (1). Moringa and fig leaves had the highest content of chlorophylls A and B but their content in moringa leaves (625.37 and 229.94 mg/100g, respectively) were more than four folds greater than fig leaves (142.14 and 83.57 mg/100g, respectively). Red pepper fruits and moringa leaves had the highest contents of total carotenoids (138.88 and 102.20 mg/100g, respectively) followed by fig leaves which were 78.83 mg/100g. These results are in agreement with those reported by Campos *et al.* (2013), Younis *et al.* (2013) and Abdalla (2015). Total anthocyanin in red pepper fruits was 25.43 mg/100g, whereas, noticeable amount was found in red pepper seeds. These results are in accordance with those reported by Loizzo *et al.* (2017).

Table 1. Bioactive compounds content of selected raw materials

Constituents	*Samples	Moringa leaves	Fig leaves	Red pepper fruits	Red pepper seeds	LSD at 5%
Anthocyanin (mg/100g)		ND	ND	25.43±0.14 ^a	1.51±1.53 ^v	0.22
Chlorophyll A (mg/100g)		625.37± 0.33 ^a	142.14± 0.39 ^p	40.77± 0.34 ^c	0.04±1.0 ^t	0.58
Chlorophyll B (mg/100g)		229.94± 1.66 ^a	83.57± 0.67 ^p	80.63±0.57 ^c	0.09±1.53 ^a	1.32
Total carotenoids (mg/100g)		102.20± 0.19 ^p	78.83± 0.48 ^c	138.88 ± 0.49 ^d	1.06±1.00 ^d	0.67
Total phenols (as gallic acid) (mg/g)		14.01± 6.42 ^a	12.12± 0.39 ^p	8.61± 0.13 ^e	4.76± 0.80 ^d	0.40
Total flavonoids (mg/g)		12.17 ± 1.26 ^a	8.02 ± 0.71 ^p	7.37± 0.36 ^q	3.82± 9.7 ^c	1.41

*(dry weight)

Moringa and fig leaves had the highest contents of total phenols and flavonoids (14.01, 12.17 and 12.12, 8.02 mg/g, respectively) followed by red pepper fruits which had 8.61 and 7.37 mg/g, respectively for total phenols and flavonoids. Meanwhile, the red pepper seeds recorded the lowest contents (4.76 and 3.82 mg/g, respectively). These results are in agreement with those reported by Shi *et al.* (2011), El Sohaimy *et al.* (2015) and Loizzo *et al.* (2017), they found that, the level of total phenolic and flavonoid contents in moringa leaves ranged from 13.4 to 115.68 and 14.32 to 113.95 mg/g, respectively, but in fig leaves ranged from 3.9 to 17.44 and 1.05 to 3.87 mg/g, respectively whereas, the total phenolic and flavonoid contents of hot red pepper varied from 1.17 to 36.87 and 0.69 to 8.39 mg/g, respectively.

Evaluation of antioxidant activity procedures of selected raw materials

Four procedures namely ABTS, DPPH, β – carotene and Frap were used to evaluate the antioxidant

activity content of moringa and fig leaves and red pepper fruits and seeds and the result are presented in Table (2). The ABTS procedure recorded the highest contents of antioxidant activity in moringa and fig leaves and red pepper fruits (93.81, 92.11 and 90.09 %, respectively) whereas; the red pepper seeds were 84.93%. Little differences of the antioxidant data between ABTS and DPPH procedures were recorded. Using DPPH procedure, the antioxidant activity of moringa, fig leaves and red pepper fruits were 89.47, 88.81 and 87.56%, respectively, while in red pepper seeds was 64.79%. These results are in accordance with those reported by Ouchemoukh *et al.* (2012), Gamal *et al.* (2016) and López *et al.* (2017).

The data of β – carotene procedure showed moderate results (68.66, 66.11, 64.40 and 61.30 %, respectively) for moringa, fig leaves, red pepper fruits and seeds comparing with the data of ABTS and DPPH procedures. Whereas, the lowest values of antioxidant activities were recorded when the Frap procedure was

used. The FRAP procedure is a method for measuring total reducing power of electron donating substances, while ABTS and DPPH assays are procedures for measuring the ability of antioxidant molecules to quench ABTS and DPPH free radicals, respectively (Wangcharoen and Gomolmanee, 2011).

Generally, from the aforementioned data it could be concluded that, the different values of each antioxidant

capacity assay resulted from the different mechanism of each procedures. The ABTS and DPPH procedures gave the highest antioxidant capacity values of all selected raw materials. Furthermore, moringa and fig leaves recorded the highest contents of antioxidant activity comparing with red pepper fruits and seeds correlated with the highly contents of total phenols and flavonoids.

Table 2. Evaluation of antioxidant activity assays of selected plants

*Samples Antioxidant activity %	Moringa leaves	Fig leaves	Red pepper fruits	Red pepper seeds	LSD at 5%
ABTS	93.81± 0.32 ^a	92.11± 0.77 ^{ab}	90.09± 2.16 ^b	84.93±1.09 ^c	2.41
DPPH	89.47±0.28 ^a	88.81±0.37 ^a	87.56±0.42 ^a	64.79±2.82 ^b	2.82
β - carotene	68.66±1.18 ^a	66.11± 0.83 ^b	64.40± 0.53 ^b	61.30±1.04 ^c	1.74
Frap	6.11± 1.17 ^a	6.00±1.53 ^b	5.79± 1.99 ^c	2.68±9.50 ^a	9.32

*(dry weight)

ABTS (+2,2-azino-bis(3-ethylbenzthiazoline- 6-sulfonic acid)).

DPPH (2, 2'-Diphenyl-1-picrylhydrazyl).

β-Carotene bleaching assay

Frap (Ferric reducing-antioxidant power).

Fractionation of phenolic compounds of selected raw materials

The phenolic compounds of selected raw materials were separated and identified by HPLC and the results are presented in Table (3). Twenty four phenolic compounds were identified and pyrogallol was the major phenolic compound detected in red pepper fruits and seeds that represented 74.19 and 54.94 mg/100g, respectively comparing with its contents in moringa and fig leaves which were 47.06 and 15.65 mg/100g, respectively. E-vanillic and vanillic acids were the most common phenolic compounds found in moringa and fig leaves (78.14 and 26.15 mg/100g, respectively), whereas, E- vanillic was the

second phenolic acid that detected in fig leaves (24.25 mg/100g), red pepper fruits (26.03 mg/100g) and red pepper seeds (18.36 mg/100g). As well as, ellagic, catechol, epicatechin and P-OH-benzoic were also detected in moderate amounts of all selected materials. These results are in agreement with those reported by Babiker *et al.* (2016) and Gamal *et al.* (2016), they stated that, the major phenolic acids was protocatechuic acid (97.92 to 109.15 mg/100g) followed by gallic acid (3.64 to 4.13 mg/100g) in moringa leaves and the most common phenolic compounds found in red pepper fruits E- vanillic acid (.15 mg/100g) followed by chlorogenic acid (15.74 mg/100g).

Table 3. Fractionation of phenolic compounds of selected plants

*Samples Phenolic compounds (mg/100g)	Moringa leaves	Fig leaves	Red pepper fruits	Red pepper seeds
Gallic acid	1.30	0.90	1.57	0.29
Pyrogallol	47.06	15.65	74.19	54.94
4-Amino-benzoic acid	0.88	0.65	1.24	2.35
Protocatechuic	4.19	2.19	4.88	4.98
Catechein	14.82	15.72	2.07	5.60
Chlorogenic acid	8.99	7.67	12.52	0.85
Catechol	11.10	8.04	27.08	3.17
Epicatechein	22.67	6.11	2.68	1.09
Caffeine	1.38	2.80	2.00	3.02
P-OH-benzoic acid	22.49	3.64	4.80	3.24
Caffeic acid	1.75	6.63	0.82	0.21
Vanillic acid	8.00	26.15	3.38	3.79
P-Coumaric acid	1.32	4.48	4.20	1.62
Ferulic acid	2.52	6.03	2.59	1.07
Iso-Ferulic acid	19.61	9.36	2.30	0.32
Reversetrol	0.42	0.79	0.42	0.32
Ellagic acid	22.00	21.95	13.37	11.29
E- vanillic acid	78.14	24.25	26.03	18.36
Alpha-Coumaric acid	7.42	3.39	2.78	0.95
Benzoic acid	17.80	19.05	4.78	8.66
3,4,5-methoxy-cinnamic acid	1.68	1.54	0.57	1.10
Coumarin	1.82	1.17	0.84	1.86
Salicylic acid	10.39	5.63	4.19	7.24
Cinnamic acid	1.65	0.69	0.27	0.26
Total	309.4	194.48	199.55	136.58

*(dry weight)

From the aforementioned data, it could be clearly concluded that moringa leaves had higher contents of phenolic acids (309.4mg/100g) than other selected plants. On the other hand, there are little differences between fig leaves and red pepper fruits but, the lowest phenolic acids content (136.58 mg/100g) was detected in red pepper seeds. These results are confirmed with the data of total

phenols content which illustrated previously in Table (1). The concentration of total polyphenols determined by Folin-Ciocalteu method was higher than the concentration obtained by HPLC method. The HPLC method presented only 50–60% of the level analyzed by the Folin-Ciocalteu method (Ferreira *et al.*, 2002).

Fractionation of flavonoid compounds of selected raw materials

Data in Table (4) reveal that, seventeen flavonoid compounds were identified from selected raw materials and hesperidin was the predominant flavonoid compound, which amounted to 55.69 mg/100g for moringa leaves, 42.64 mg/100g for red pepper fruits and 30.52 mg/100g for seeds. Rutin was the major compound (75.21 mg/100g) found in fig leaves, but it was the second major flavonoid compound (48.75 mg/100g) detected in moringa leaves. Furthermore, hesperidin (47.53 mg/100g) in fig leaves and naringenin (20.76 mg/100g and 20.09 mg/100g) in red pepper fruits and seeds were the second major flavonoids

compounds detected. These results are in accordance with those reported by Gamal *et al.* (2016) who found that fig leaves were rich sources for many flavones as Kampherol and rutin. Moreover, apigenin -7 -glucose, acacetin neo. rutinodie and luteolin found also in high amounts in all selected raw materials.

Generally, it could be clearly concluded that there were variation in flavonoids individuals between all selected raw materials. Moringa and fig leaves had approximately the same highest contents of flavonoids (283.71 and 266.73 mg/100g, respectively) than red pepper seeds which recorded lowest ones (112.84 mg/100g).

Table 4. Identification of flavonoid compounds of selected plants

Flavonoid compounds (mg/100g)	*Samples Moringa leaves	Fig leaves	Red pepper fruits	Red pepper seeds
Luteolin	17.44	20.32	6.18	15.32
Naringenin	44.60	32.22	20.76	20.09
Rutin	48.75	75.21	9.162	3.07
Hesperidin	55.69	47.53	42.64	30.52
Rosmarinic	4.49	2.00	0.92	1.27
Quercetrin	4.85	6.67	7.28	2.19
Quercetin	3.31	1.37	1.49	1.82
Naringenin	8.05	1.46	2.54	0.32
Hesperidin	11.31	2.20	4.43	0.92
Kaempferol	2.77	0.42	2.06	1.41
Apigenin	1.14	0.55	1.12	1.63
Apigenin 6- rhamose 8- glucose	9.51	17.76	6.82	14.80
Quercetin-3-o-glucose	0.69	2.90	1.08	0.097
Apigenin -7 -glucose	32.47	33.64	15.91	2.55
Apigenin -7-0 -neoheos	4.76	7.19	10.08	5.65
Kaempferol 3-7-diramoside	9.63	4.31	3.96	2.74
Acacetin neo. rutinodie	24.25	10.62	17.68	8.24
Total	283.71	266.37	151.11	112.84

*(dry weight)

B complex vitamins content of selected raw materials

Data in Table (5) reveal that, cobalamin (B₁₂) was the major B complex vitamins found in fig leaves, red pepper fruits, moringa leaves and red pepper seeds which recorded 994.01, 217.46, 164.81 and 125.5 mg/100g, respectively. Whereas, nicotinic acid was the second detected vitamins in red pepper fruits, fig leaves and red pepper seeds (70.22, 38.84 and 30.81 mg/100g, respectively) as well as, pyroxidin (B₆) was 70.09 mg/100g in moringa leaves. Thiamin (B₁), riboflavin (B₂) and folic acid detected also with small amounts in all tested plants. The aforementioned data of B complex vitamins are in accordance with those reported by Mensah *et al.* (2012) and El Sohaimy *et al.* (2015), they reported that, the vitamin B complex content of moringa leaves ranged from 0.05 to 2.64 (B₁) and 0.8 to 20.5(B₂).

Vitamins A, C, D, E and K contents of selected raw materials

From the data in Table (6), it could be clearly noticed that moringa leaves and red pepper fruits had the highest content of vitamins K (725.25 and 530.90 mg/100g, respectively) and vitamin C (317.72 and 118.53

mg /100g), followed by fig leaves and red pepper seeds which contains moderate amounts. These results are in agreement with those reported by Keleş *et al.* (2016) and Nobosse *et al.* (2017) they found that, moringa leaves and red pepper fruits are rich sources of vitamin C which ranged from 218.13 to 691.0 and 10.10 to 100.5 mg/100g, respectively, as well as, peppers had ascorbic acid content four times higher than orange that are considered the most important source for this healthy compound. The vitamin C content of fresh and dried hot red pepper ranged from 87.13 to 280.0 and 58.5 to 173.7 mg/100g, respectively (Ozgun *et al.*, 2011). On the other hand, small amounts of vitamin D and noticeable amounts of vitamins A and E were detected in all tested raw materials.

Generally, it could be concluded that moringa leaves and red pepper fruits are considered as excellent sources for vitamins C and K that play an important roles as antioxidant agents to protect biological systems via inhibition or prevention of oxidation stress induced by reactive oxygen substances generated from normal metabolic activity or environmental factors followed by red pepper seeds and fig leaves.

Table 5. Fractionation of B complex vitamins of selected plants

Vitamin B Complex fractions (mg/100g)	*Samples Moringa leaves	Fig leaves	Red pepper Fruits	Red pepper seeds
Nicotenic acid (Niacin)	39.25	38.84	70.22	30.81
Thiamin (B1)	19.81	8.93	19.36	6.25
Pyroxidin (B6)	70.09	10.47	22.79	7.95
Folic acid	8.65	6.06	12.35	5.63
Cobalamin (B12)	164.81	994.01	217.46	125.5
Riboflavin (B2)	16.25	13.12	11.55	4.32

*(dry weight)

Table 6. Vitamins A, C, D, E and K contents of selected plants

Vitamins (mg/100g)	*Samples	Moringa leaves	Fig leaves	Red pepper fruits	Red pepper seeds
Vit. A		0.37	1.01	0.79	0.31
Vit. C		317.72	41.95	118.53	23.08
Vit. D		3.21	6.38	5.81	2.85
Vit. E		0.42	0.28	0.16	0.1
Vit. K		725.25	86.45	530.90	211.45

*(dry weight)

REFERENCES

- Abdalla, Mona, M. (2015). The potential of *Moringa oleifera* extract as a biostimulant in enhancing the growth, biochemical and hormonal contents in rocket (*Eruca vesicaria subsp. sativa*) plants. Afr. J. Crop Sci., 3(2): 116-122.
- Anjorin, T.; Ikokoh, P. and Okolo, S. (2010). Mineral composition of *Moringa oleifera* Leaves, pods and seeds from two regions in Abuja, Nigeria. Int. J. Agric. Biol., 12: 431-434.
- Babiker, E.; Juhaimi, F.; Ghafoor, K. and Abdoun, K. (2016). Effect of drying methods on nutritional quality of young shoots and leaves of two Moringa species as non-conventional fodders. Agro forest Syst., DOI 10.1007/s10457-016-0043-8.
- Batifoulie, F.; Verny, M.; Besson, C.; Demigne, C. and Remesy, C. (2005). Determination of thiamin and its phosphate esters in rat tissues analyzed as thiochromes on RP- amide C16 column. J. Chrom. B., 8(16): 67-72.
- Brand-Williams, W.; Cuvelier, M. and Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. Lebensm. Wiss. Tech., 28: 25-30.
- Campos, M.R.; Gómez, K. R.; Ordoñez, Y.M. and Ancona, D. B. (2013). Polyphenols, ascorbic acid and carotenoids contents and antioxidant properties of Habanero pepper (*Capsicum chinense*) fruit. Food and Nutrition Sciences, (4):47-54.
- Dapkevicius, A.; Venskutonis, R.; Van Beek, T. and Linsen, P. (1998). Antioxidant activity of extracts obtained by different isolation procedures from some aromatic herbs grown in Lithuania. J. Sci. Food Agric., 77:140-145.
- El Sohaimy, S.; Hamad, G.; Mohamed, S.; Amar, M. and Al-Hindi, R. (2015). Biochemical and functional properties of *Moringa oleifera* leaves and their potential as a functional food. Global Advanced Research Journal of Agricultural Science, 4(4): 188 – 199.
- Ferreira, D.; Guyot, S.; Marnet, N.; Delgadillo, I.; Renard, C.M. and Coimbra, M.A., (2002). Composition of phenolic compounds in a portugese pear (*Pyrus communis L. Var. S. Bartolomeu*) and changes after sun drying. J. Agric. and Food Chem., 50: 4537-4544.
- Gamal, S.A.; Mahmoud, M. S.; Mohamed, M. R. and Emam, A. A. (2016). Phytochemical screening, antioxidant activities and in vitro anticancer potential of Egyptian Capsicum Spp. Biochem Pharmacol. (Los Angel), 5(2).
- Gilani, A.; Mehmood, M.; Janbaz, K.; Khan, A. and Saeed, S. (2008). Ethnopharmacological studies on antispasmodic and antiplatelet activities of *Ficus carica*. J Ethno pharmacol., 119: 1-5.
- Goupy, P.; Hugues, M.; Biovin, P. and Amiot, M.J. (1999). Antioxidant composition and activity of barley (*Hordeum vulgare*) and malt extracts and isolated phenolic compounds. J. Sci. Food Agric., 79: 1625-1634.
- Herre, E.; Jander, K. and Machado, C. (2008). Evolutionary ecology of figs and their associates: recent progress and outstanding puzzles. Ann Rev Ecol. Evol. Syst., 39:439-458.
- Holm, G. (1954). Chlorophyll mutations in barley. Acta. Agr. Scand., 4: 457-471.
- Jia, Z.; Tang, M. and Wu, J. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. Food Chem., 64: 555-559.
- Lim, Y. Y. and Murtijaya, J. (2007). Antioxidant properties of *Phyllanthus amarus* extracts as affected by different drying methods. LWT – Food Sci. Tech., 40: 1664-1669.
- Loon, Y.; Wong, J.; Yap, S. and Yuen, K.(2005). Determination of flavonoids from Orthosiphon stamineus in plasma using a simple HPLC method with ultraviolet detection. J. Chromatogr. B., 816: 161- 166.
- López, R. I.; Félix, J. L.; Escalante, M. A.; Dorado, R. G.; Rangel, M. D. and Heredia, J. B. (2017). Nutritional and phenolic characterization of *moringa oleifera* leaves grown in Sinaloa, Mexico. Pak. J. Bot., 49(1): 161-168.
- Karimi, E.; Jaafar, H. and Ahmad, S. (2011). Phenolics and flavonoids profiling and antioxidant activity of three varieties of Malaysian indigenous medicinal herb *Labisia pumila* Benth. J. Med. Plants Res., 5: 1200-1206.
- Keleş, D.; Özgen, S.; Saraçoğlu, O.; Ata, A. and Özgen, M. (2016). Antioxidant potential of Turkish pepper (*Capsicum annum L.*) genotypes at two different maturity stages. Turk J Agric For., 40: 542-551. doi:10.3906/tar-1601-24.
- Kim, G. D.; Lee, Y. S.; Cho, J.-Y.; Lee, Y. H.; Choi, K. J.; Lee, Y. and *et al.* (2010). Comparison of the content of bioactive substances and the inhibitory effects against rat plasma oxidation of conventional and organic hot peppers (*Capsicum annum L.*). J Agric Food Chem., 58, 12300-12306.
- Mahajan, S. G.; Banerjee, A.; Chauhan, B. F.; Padh, H.; Nivsarkar, M. and Mehta, A. A. (2009). Inhibitory effect of n-butanol fraction of *Moringa oleifera* Lam. seeds on ovalbumin-induced airway inflammation in a guinea pig model of asthma. Int. J. Toxicol., 28(6): 519-527. DOI: 10.1177/1091581809345165.
- Maurya, S. and Singh, D. (2010). Quantitative analysis of total phenolic content in *Adhatoda vasica* nees extracts. Int J. Pharm. Tech. Res., 2(4): 2403-2406.

- Mehta, K.; Balaraman, R.; Amin, A.H.; Bafna, P.A. and Gulati, O.D. (2003). Effect of fruits of *Moringa oleifera* on the lipid profile of normal and hypercholesterolaemic rabbits. J. Ethnopharmacol., 86(2-3):191-195.
- Mensah, J.; Ikhajagbe, B.; Edema, N. and Emokor, J. (2012). Phytochemical, nutritional and antibacterial properties of dried leaf powder of *Moringa oleifera* (Lam) from Edo Central Province, Nigeria. J. Nat. Prod. Plant Resour., 2(1): 107-112.
- Nishino, H.; Murakoshi, M.; Tokuda, H. and Satomi, Y. (2009). Cancer prevention by carotenoids. Arch Biochem Biophys., 483:165-168.
- Ouchemoukh, S.; Hachoud, S.; Boudraham, H. and Mokrani, A. (2012). Antioxidant activities of some dried fruits consumed in Algeria. Food Sci. Technol., 9: 329-332.
- Perez, C.; Canal, J.R. and Torres, M.D. (2003). Experimental diabetes treated with *Ficus carica* extract: effect on oxidative stress parameters, Acta Diabetologica., 40:3-8.
- Pyka, A. and Sliwiok, J. (2001). Chromatographic separation of tocopherols. J. Chromatogr. A., 935(1-2):71-76.
- Rahman, M.M. and Sheikh, M.M.I. (2009). Antibacterial activity of leaf juice and extracts of *Moringa oleifera* Lam. against some human pathogenic bacteria. CMU. J Nat Sci., 8(2):225.
- Rangana, S. (1977). Fruit and Vegetable Analysis. Manual of analysis of fruit and vegetable products. Tata. Mc. Graw- Hill, Pub. Co. Ltd, New Delhi.
- Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radic Biol Med., 26: 1231-1237.
- Shi, Y. X.; Xu, Y. K.; Hu, H. B.; Na, Z. and Wang, WH. (2011). Preliminary assessment of antioxidant activity of young edible leaves in Xishuangbanna, Southwest China. Food Chem., Doi: 10.1016/J.Food chem.03.113.
- Siddhuraju, P. and Becker, K. (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves. J Agric Food Chem., 51: 2144-2155.
- Steel, R.; Torrie, J. and Dickey, D. (1997). Principles and procedures of Statistics: A Biometrical Approach, 3rd ed., McGraw-Hill, New York, NY.
- Stoys, S. and Hartman, M. (2015). Review of the safety and efficacy of *Moringa oleifera*. Phytother Res., 29(6): 796-804.
- Tunde-Akintunde, T. (2010). Effect of pretreatment on drying time and quality of chilli pepper. J. Food Process. Preserv., 34: 595-608.
- Volko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.; Mazur, M. and Telsler, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. Int. J. Biochem. Cell Bio., 39: 44-84.
- Wahba, N.; Ahmed, A. and Ebraheim, Z. (2010). Antimicrobial Effects of Pepper, Parsley, and Dill and Their Roles in the Microbiological Quality Enhancement of Traditional Egyptian Kareish Cheese. Foodborne Pathog. Dis., 7: 411-418.
- Wangcharoen, W. and Gomolmanee, S. (2011). Antioxidant Capacity and Total Phenolic content of *Moringa oleifera* grown in Chiang Mai, Thailand. Thai J. Agric. Sci., 44(5): 118-124.
- Wettstein, D. (1957). Chlorophyll – letale und der submikroskopische ormwechsel der Plastiden. Exp. Cell Res., 12, 427-487.
- Younis, U.; Bokhari, T.; Raza Shah, M.; Mohmood, S. and Malik, S. (2013). Dust interception capacity and alteration of various biometric and biochemical attributes in cultivated population of *ficus carica* L. Journal of Pharmacy and Biological Sciences., 6(4): 35-42.

دراسة مقارنة على بعض النباتات كمصادر لمضادات الأكسدة الطبيعية إيمان محمد صديق الأشعل¹، أحمد عبد العزيز الرفاعي² وأمل محمد البسطويسى³ ¹معهد بحوث تكنولوجيا الأغذية – مركز البحوث الزراعية – الجيزة – مصر. ²قسم الصناعات الغذائية – كلية الزراعة – جامعة المنصورة – مصر.

في الأونة الأخيرة ظهر اهتمام عالمي كبير بالنباتات والمنتجات الطبيعية نتيجة لدخولها في العديد من التطبيقات الغذائية والدوائية ومستحضرات التجميل . لذلك تم تقييم المركبات الحيوية النشطة لكل من أوراق نبات المورينجا والتين وثمار وبذور الفلفل الأحمر كمضادات طبيعية للأكسدة بالإضافة إلى تقييم النشاط المضاد لإكسده لهذه النباتات باستخدام طرق مختلفة والمفاضله بينها. وتوصلت النتائج إلى أن أوراق نبات المورينجا والتين سجلت أعلى محتوى من كلورفيل A & B حيث كان محتواها في أوراق المورينجا (٦٢٥.٣٧ و ٢٢٩.٩٤ ملجرام/١٠٠ جرام على التوالي) أعلى أربع أضعاف من محتواها في أوراق التين والتي كانت ١٤٢.١٤ و ٨٣.٥٧ ملجرام /١٠٠ جرام على التوالي. وأحتوت ثمار الفلفل الأحمر وأوراق المورينجا على أعلى نسبة من الكاروتينبات الكلية. كما سجلت كلا من طريقه ABTS و DPPH أعلى قيم في النشاط المضاد للأكسده في جميع العينات مقارنة بالطرق الأخرى. وسجلت أوراق المورينجا والتين أعلى محتوى من النشاط المضاد للأكسده مقارنة بثمار وبذور الفلفل وذلك نتيجة لإرتفاع محتواها من الفينولات والفلافونيدات الكلية. سجلت أوراق المورينجا أعلى محتوى من الأحماض الفينولية (٣٠٩.٤ ملجرام/١٠٠ جرام) مقارنة بباقي العينات وكان E-vanillic و Pyrogallol أكثر المركبات الفينولية شيوعا في أوراق نبات المورينجا كما وجد إختلافات طفيفة بين أوراق التين وثمار الفلفل الأحمر بينما سجلت بذور الفلفل الأحمر أقل محتوى من المركبات الفينولية (١٣٦.٥٨ ملجرام /١٠٠ جرام). كما وجد إختلافات في محتوى الفلافونيدات المفردة بين كل المواد محل الدراسة حيث سجلت أوراق المورينجا والتين تقريبا نفس المحتوى المرتفع في الفلافونيدات (٢٨٣.٧١ و ٢٦٦.٧٣ ملجرام /١٠٠ جرام) مقارنة بثمار الفلفل الأحمر وبذوره والتي كانتا أقلهما في محتوى الفلافونيدات (١١٢.٨٤ ملجرام/١٠٠ جرام). كان الكوبلامين (B₁₂) من أكثر مجموعة فيتامينات B المركبة التي وجدت في أوراق التين وثمار الفلفل الأحمر وأوراق المورينجا وبذور الفلفل الأحمر والتي سجل ٩٩٤.٠١ و ٢١٧.٤٦ و ١٦٤.٨١ و ١٢٥.٥ ملجرام/١٠٠ جرام على التوالي. وتعتبر أوراق نبات المورينجا وثمار الفلفل الأحمر من المصادر الممتازة لفيتامين C و K والتي تلعب دور مهم كمضادات أكسده لحماية النظام الحيوى وتنشيط ومنع الجهد التأكسدى الناتج عن جزيئات الأوكسجين النشطة الناتجة من نشاط الميتابولزم الطبيعي للجسم أو العوامل البيئية الخارجية ويلبيها بذور الفلفل الأحمر وأوراق التين.