

## Effect of Thyme and Rosemary on Liver Functions in Toxicated Rats by Aflatoxins

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### ABSTRACT

Thyme and rosemary assume a part in hepatoprotectivity and go about as anti-aflatoxicosis. This examination has endeavored to explore impact of subacute measurements of aflatoxin in rats and conceivably to foresee any advantages or disadvantages of thyme and rosemary. Thirty adult male albino rats were partitioned into five groups. Rats were isolated into two main groups, the first main group (n=6 rats), negative control, was fed on basal diet. The second main group was inebriated with Aflatoxin then was divided into four subgroups as follow, the first subgroup was kept as positive control group, sub groups (2 and 3) were fed on basal diet and supplemented with thyme and rosemary at of 5% individually. The fourth subgroup was fed on basal diet and supplemented with a combination of 2.5% thyme and 2.5% rosemary. The results indicated that, supplementation with Thyme, Rosemary and their mixes altogether significantly ( $P<0.05$ ) enhanced liver functions and also serum protein parameters (albumin, globulin and total protein), serum CAT, SOD and decrease significantly MDA compared to the positive control. Furthermore Thyme, Rosemary and their blend significantly decreased ( $P<0.05$ ) the mean level of serum TNF- $\alpha$  and IL-1- $\beta$  contrasted with the positive control gathering. The present examination prescribed utilizing Thyme, Rosemary and their blends to shield the body from aflatoxin danger.

**Keywords:** Thyme, Rosemary, Aflatoxin, Liver functions, Phenolic, antioxidant Rats.

### INTRODUCTION

Aflatoxins are the most imperative mycotoxins, perceived as universal contaminants of sustenance all through developing world (Kamkar *et al.*, 2013). Samuel *et al.*, (2013) demonstrated that, the significant aflatoxins are AFB1, AFB2, AFG1, AFG2. Among them, aflatoxin B1 (AFB1) is the most strong reason for human cancer-causing agent (Reddy *et al.*, 2009b and Tavakoli *et al.*, 2013).

Many studies by (Peters and Teel, 2003; Gong *et al.*, 2004; Williams *et al.*, 2004 and Abnet, 2007) revealed that aflatoxins are created by a few types of the fungus *Aspergillus*. Aflatoxicosis can cause intermittent genuine medical issues and important economic losses. No less than 100 nations have directions to control significant mycotoxins, particularly aflatoxins, in products and nourishment, with the goal that the most extreme bearable mycotoxins levels fluctuate incredibly among the countries (Reddy *et al.*, 2009a).

Oxidative pressure, including lipid peroxidation, has the significant job in the pathogenesis of aflatoxicosis (Abdel-Wahhab *et al.*, 2006 and Umarani *et al.*, 2008). Chemoprevention of toxicoses utilizing supplements is the subject of exceptional investigation. Antioxidants have the ability to reduce disease formation by either enlistment or restraint of key catalyst frameworks (Guarisco *et al.*, 2008).

Herbs are normally wealthy in bioactive plant items with nourishment esteem as considerable restorative incentive in a few illnesses (Sharma, 2010). *Thyme* (*Thymus vulgaris*) was having a place with the *Lamiaceae* family a sweet-smelling local herb in the Mediterranean area. Thyme was presently broadly developed as herbal medicine (Domaracky *et al.*, 2007). *Thymus vulgaris* possess various beneficial effects, like antiseptic, antimicrobial, bactericidal and antioxidant properties. Also, it has recommended as a natural replacement for synthetic antioxidant (Rasooli *et al.*, 2006). Höferl *et al.*, (2006) found that, thyme functions as a liver decontamination tonic, promotes blood circulation and functions as an exciting stimulant for the entire system. The therapeutic

potential of thymus is due to its contents of flavonoids, thymol, carvacrol, eugenol and aliphatic phenols in addition to luteolin and saponins (Amarowicz *et al.*, 2009). These flavonoids advance ideal wellbeing through their antioxidant capacity and rummaging free radicals (El-Nekeety, 2011). Aqueous extract of thyme was wealthy in the phenolic content and have free radical searching movement (Hamzawy *et al.*, 2012). Thymol and carvacrol are recognizable antioxidants found in the concentrate of thyme species plants (Beena and Rawat, 2013).

Rosemary (*Rosemarinus officinalis L.*) an evergreen perpetual fragrant bush having a place with the family *Labiatae* (Al-Sereiti *et al.*, 1999). It is normally utilized as a zest and enhancing operator (Saito *et al.*, 2004). It is made out of dried leaves and blooms contain some antioxidant (flavonoids and phenols) which thought about having antioxidant properties (Nabavi *et al.*, 2015). These polyphenols have demonstrated natural activities in vitro as hostile to tumor, chemopreventive (Razavi-Azarkhiavi *et al.*, 2014) and anti-inflammatory agents and may assume a job by directing the movement and additionally articulation of certain enzymatic frameworks ensnared in important physiological procedures in the liver (Del Bano *et al.*, 2006).

The antioxidant activity movement of rosemary think can be attributed basically to two sections, carnosic destructive and carnosic Kadri *et al.*, 2011 and Machado *et al.*, 2013).

**Aim of study:** The purpose behind the present work was to survey the effect of Thyme and Rosemary supplementation on liver function and oxidative worry in toxicated rats by Aflatoxins.

### MATERIALS AND METHODS

#### Materials:

**Rats:** Thirty adult male albino rats were obtained from Helwan Farm, Ministry of Health and Population, Cairo, Egypt. Diet: Casein, vitamins, cellulose, minerals, methionine and choline were obtained from Morgan Company for Chemicals, Cairo, Egypt. Plants: Leaves of Rosemary (*Rosmarinus officinalis*)

and thyme (*Thymus vulgaris* L.) were obtained from the Agriculture Faculty, Cairo University and was identified by competent botanist at the herbarium of Agriculture Research Center, Giza, Egypt. Chemicals: Chemical kits were purchased from Gama Trade Company for Chemicals, Cairo, Egypt. Purified AFB1 from *Aspergillus flavus* was purchased from Sigma Company, Cairo, Egypt.

#### Methods:

**Experimental design:** In this study 30 rats, weighing (180 ± 10 g) were kept under normal healthy condition and fed on basal diet for one week for adaptation (Reeves *et al.*, 1993). Rats were divided into two main groups, the first main group (n=6 rats), negative control, was fed on basal diet. The second main group was intoxicated with Aflatoxin, (Aflatoxin B1) through intragastrically route at concentrations 2 mg /kg b.w., for 7 days) according to (Wójtowicz-chomicz *et al.*, 2011) then were divided into four subgroups as follow, the first subgroup was kept as positive control group, subgroups (2 and 3) were fed on basal diet and supplemented with thyme and rosemary at the level of 5% respectively. The fourth subgroup was fed on basal diet and supplemented with a combination of 2.5% thyme and 2.5% rosemary. After 8 weeks rats were sacrificed after overnight fasting and blood of each rat was taken from the vein of the eye. The serum was separated at 3000 rpm for 20 minutes by centrifuge, then kept in plastic vials at -20°C until analysis.

#### Biochemical Analysis:

Serum liver enzymes including aspartat aminotransaminase (AST), alanine aminotransaminase (ALT) and alkaline phosphatase (ALP) were determined (Reitman and Frankel, 1957). Serum total protein, albumin and globulin were estimated according to Weissman *et al.*, (1950). Oxidative stress markers: serum Catalase (CAT), Superoxide Dismutase (SOD) and malondialdehyde (MDA) were determined according to (Beutler *et al.*, 1963; Kakkar *et al.*, 1984 and Draper and Hadly, 1990) respectively.

#### Statistical analysis:

The obtained data was statistically analyzed using the Statistical Package for Social Science (SPSS) version 18.0. Values are represented as means with their standard errors (SE). P value of < 0.05 was considered to indicate statistical significance (Snedecor and Cochran, 1980).

## RESULTS

Table (1) showed the effect of Thyme, Rosemary and their mixture on serum liver functions in rats with induced toxicity by Aflatoxins. Results indicated that the positive control group had a significant increase (P<0.05) in serum levels of AST, ALT and ALP compared to the healthy group. Supplementation with Thyme, Rosemary and their mixture at the tested level significantly decreased (P<0.05) the elevated levels of serum ALT, AST and ALP compared to the positive control group. Moreover, there was significant difference in serum ALT, AST and ALP between thyme and rosemary treated groups. The best results of liver functions were recorded at the group fed on 2.5% of both Thyme and Rosemary.

**Table 1. Effect of Thyme, Rosemary and their mixture on liver functions of intoxicated rats by Aflatoxins.**

Parameters Groups	AST	ALT (μ/L)	ALP
Control (-ve)	90.30±2.40 <sup>c</sup>	27.40±1.07 <sup>d</sup>	105.63±3.53 <sup>e</sup>
Control (+ve)	162.16±4.64 <sup>a</sup>	61.66±1.81 <sup>a</sup>	199.36±3.27 <sup>a</sup>
Thyme (5%)	116.36±1.19 <sup>c</sup>	34.13±1.69 <sup>c</sup>	147.16±1.81 <sup>c</sup>
Rosemary (5%)	127.03±1.76 <sup>b</sup>	39.67±1.45 <sup>b</sup>	161.93±3.12 <sup>b</sup>
Mix. (2.5% Thyme + 2.5% Rosemary)	105.60±3.00 <sup>d</sup>	32.73±0.89 <sup>c</sup>	133.50±2.56 <sup>d</sup>

Values are expressed as means ± SE.

Values at the same column with different letters are significantly different at P<0.05.

Data in table (2) illustrated the effect of Thyme, Rosemary and their mixture on serum (albumin, globulin and total protein) of intoxicated rats by Aflatoxins. It could be observed that, the positive control rats had significant reduction (P<0.05) in the mean value of serum albumin, globulin and total protein compared to the negative control group. The supplementation with Thyme, Rosemary and their mixture significantly increased (P<0.05) the mean level of serum protein parameters compared to the positive control group. There was significant difference in serum protein parameters between thyme and rosemary treated groups. The concentration of serum protein parameters significantly increased (P<0.05) as a results of supplementation with 2.5% of both Thyme and Rosemary. Moreover, there was no significant change in serum albumin and globulin between the two treated groups (Thyme and their mixture).

**Table 2. Effect of Thyme, Rosemary and their mixture on serum protein parameters of intoxicated rats by Aflatoxins**

Parameters Groups	Albumin	Globulin (g/dl)	Total Protein
Control (-ve)	5.76±0.24 <sup>a</sup>	3.13±0.14 <sup>a</sup>	9.80±0.23 <sup>a</sup>
Control (+ve)	2.72±0.17 <sup>d</sup>	1.40±0.09 <sup>d</sup>	4.30±0.15 <sup>e</sup>
Thyme 5%	4.63±0.08 <sup>b</sup>	2.38±0.12 <sup>b</sup>	6.73±0.27 <sup>c</sup>
Rosemary 5%	3.93±0.17 <sup>c</sup>	1.94±0.04 <sup>c</sup>	5.80±0.41 <sup>d</sup>
Mix. (2.5% Thyme+ 2.5% Rosemary)	5.03±0.20 <sup>b</sup>	2.65±0.05 <sup>b</sup>	7.90±0.26 <sup>b</sup>

Values are expressed as means ± SE.

Values at the same column with different letters are significantly different at P<0.05.

Table (3) revealed the effect of Thyme, Rosemary and their mixture on serum CAT, SOD and MDA of intoxicated rats by Aflatoxins. Injection with Aflatoxins significantly decreased (P<0.05) the mean value of serum CAT and SOD but caused an increase in the level of MDA compared to the negative control group. The supplementation with Thyme, Rosemary and their mixture significantly increased (P<0.05) the mean level of serum CAT and SOD and decrease significantly serum MDA compared to the positive control group. There was no significant difference in serum CAT and SOD between thyme and rosemary tested groups. The best results for the concentrations of CAT, SOD and MDA were recorded at the group fed on basal diet supplemented with 2.5% combination of (Thyme and Rosemary).

**Table 3. Effects of Thyme, Rosemary and their mixture on serum Catalase, Superoxide Dismutase and malondialdehyde of intoxicated rats by Aflatoxins**

Parameters Groups	CAT (μ/L)	SOD (μ /dl)	MDA (ng/ml)
Control (-ve)	100.10±2.62 <sup>a</sup>	95.13±2.67 <sup>a</sup>	8.06±0.70 <sup>d</sup>
Control (+ve)	24.20±2.95 <sup>e</sup>	48.10±2.12 <sup>e</sup>	36.83±2.47 <sup>a</sup>
Thyme 5%	60.23±3.09 <sup>c</sup>	75.20±1.56 <sup>c</sup>	23.46±1.89 <sup>b</sup>
Rosemary 5%	47.10±1.81 <sup>d</sup>	63.76±2.47 <sup>d</sup>	28.40±2.21 <sup>b</sup>
Mix. (2.5% Thyme + 2.5% Rosemary)	76.00±2.64 <sup>b</sup>	83.83±2.34 <sup>b</sup>	15.20±2.16 <sup>c</sup>

Values are expressed as means ± SE.

Values at the same column with different letters are significantly different at P<0.05.

The effect of Thyme, Rosemary and their mixture on serum Tumor Necrosis Factor and interleukin-1 intoxicated rats by Aflatoxins were recorded in Table (4). The positive control rats had significant increased (P<0.05) in the mean value of serum TNF-α and IL-1-β, compared to the negative control group. Thyme, Rosemary and their mixture significantly decreased (P<0.05) the mean level of serum TNF-α and IL-1-β compared to the positive control group. There was significant difference in serum TNF-α and IL-1-β among the thyme and rosemary treated groups. Supplementation with 2.5% combination of Thyme and Rosemary gave the best results of serum TNF-α and IL-1-β.

**Table 4. Effects of Thyme, Rosemary and their mixture on serum Tumor Necrosis Factor and interleukin-1 of intoxicated rats by Aflatoxins**

Parameters Groups	TNF-α (pg/ml)	IL-1β (pg/ml)
Control (-ve)	55.26±2.85 <sup>e</sup>	0.67±0.01 <sup>d</sup>
Control (+ve)	132.26±1.89 <sup>a</sup>	2.17±0.13 <sup>a</sup>
Thyme 5%	100.76±3.03 <sup>c</sup>	1.20±0.05 <sup>c</sup>
Rosemary 5%	118.23±1.68 <sup>b</sup>	1.50±0.11 <sup>b</sup>
Mix. (2.5% Thyme+ 2.5% Rosemary)	92.50±1.32 <sup>d</sup>	0.95±0.02 <sup>c</sup>

Values are expressed as means ± SE.

Values at the same column with different letters are significantly different at P<0.05.

## DISCUSSION

Liver is the main organ to experience ingested supplements, drugs and ecological toxicants that enter the hepatic entry vein from the stomach related framework and liver capacity can be changed by damage coming about because of intense or endless presentation to toxicants. Aflatoxin B1 is a hepatotoxic and hepatocarcinogenic compound delivered by the growth, *Aspergillus (A.) flavus* (Eaton and Gallagher, 1994). An assortment of nourishments, for example, grains, millets and oil seeds, are powerless to contamination by *A. flavus*, which produces aflatoxins amid its development, reap, transport and capacity. AFB1 is likewise bio changed by P450 catalysts to yield an electrophilic epoxide (Kodama *et al.*, 1990), which assaults the DNA to start hepatotoxicity (Shen *et al.*, 1996).

Low patient satisfaction from the consumption of synthetic drugs, due to high costs and side effects of these medications caused an increased tendency to traditional treatments (Al-Attar and Shawush, 2015). Extraordinary consideration has been paid to the defensive impacts of antioxidants by regular root mixes against harming caused

by concoction operators (Abdel-Wahhab *et al.*, 2011). Phenolic mixes with antioxidant activities have been appeared to have defensive impacts on different organs (Amarowicz *et al.*, 2009).

Fresh Thyme herb has a standout amongst the most antioxidant levels among herbs. It is squeezed with minerals and vitamins that are fundamental for perfect prosperity. Its leaves are affluent in potassium, squeeze, calcium, manganese, magnesium and selenium (Sharangi and Guha, 2013). Rosemary and its constituents especially caffeic corrosive subordinates, for example, rosmarinic corrosive have a remedial potential in anticipation of inflammatory diseases and hepatotoxicity (Al-Sereiti *et al.*, 1999).

The liver functions were analyzed through the assurance of ALT and AST activities which known as cytosolic marker catalysts reflecting hepatocellular putrefaction as they are discharged into the blood after cell layer harm damage (Andallu and Vardacharyulu, 2001). The present outcomes demonstrate a significant increase in AST, ALT and ALP because of aflatoxin infusion. Rise of liver catalysts was as per that revealed by many examinations (Salama *et al.*, 2013; Zargar, 2014 and Kim *et al.*, 2014). Sherif *et al.*, (2009) indicated that liver enzymes increased after liver damage because of increased membrane permeability or because of liver cell necrosis and cytosol leakage into the serum.

The treatment of rats with Thyme, Rosemary and their combination improved the liver functions that decreased by AFB1 intoxication as seen in table (1). Shanon, (2011) reported that adding aqueous extract of thyme at 10% to drinking water for the mothers of broiler chickens causes decline the level of liver enzymes, this may be due to thyme compounds that enhance the status of antioxidants. Thyme contains flavonoids that due to decline in level of ALP by obstruction operation in the liver and bones (Wathfeq *et al.*, (2011).

Likewise past investigations concur with the consequences of the present examination (Al-Kattan *et al.*, 2012). Thymol stifled cytochrome P450 intervened metabolic enactment of CCl4 (Al-Malki, 2010). Moreover, carvacrol applied antioxidant and hepatoprotective impacts in rats (Aristatile *et al.*, 2009).

Rosemary, through one of its dynamic constituent's rosmarinic corrosive, is regularly utilized for its advantageous impacts on the liver infection (Li *et al.*, 2010a). Abdel-Wahhab *et al.*, (2011) revealed that organization of rosemary extract previously or amid the treatment with CCl4 enhanced every biochemical parameter and histological photo of the liver by searching or hindering the arrangement of free radicals created amid CCl4 digestion. Awad *et al.*, (2011) detected that the treatment of aflatoxicated rats with extract of rosemary resulted in a significant degradation of aflatoxins from liver and kidney. The ameliorative effect of rosemary may be due to its antioxidant properties in combating free radical-induced oxidative stress and tissue injury (Sakr *et al.*, 2015).

The current study revealed that the supplementation with Thyme, Rosemary and their mixture significantly increased (P<0.05) the mean level of serum protein parameters compared to the positive control group. Similar

results were obtained by (Haroun *et al.*, 2002 and Mansour *et al.*, 2002).

Seung *et al.*, (2005) detailed that thyme have a successful defensive instrument because of responsive oxygen species and might be related with diminished the oxidative stress. As reported by Abd El Kader and Mohamed, (2012) it is feasible for the thyme extract to be mediated its antioxidant activities by enhancing the antioxidant defense enzymes SOD, CAT and replenishing GSH storage. Furthermore, thyme extract which show antioxidant activity has an inhibitory effect on lipid peroxidation, which could decrease the strength of inflammatory response (Bozin *et al.*, 2006). The same results were obtained by Rubió *et al.*, (2014).

Rosemary enhanced the toxic impacts of AFB1 on liver. This was showed by decrease of the level of MDA and increment CAT and SOD. The obtained results about oxidative stress are in concurrence with Parmar *et al.*, (2011), who reported that treatment with aqueous extract of rosemary leaves prevents oxidative stress due to carnosic corrosive (CA), carnosol and other phenolic acids in rosemary (Abd El-Ghany *et al.*, 2012). CA can avert lipid peroxidation and it avoids the disruption of the biological membrane by searching free radicals (Munné-Bosch and Alegre, 2001). Virk *et al.*, (2013) reported also that rosemary aqueous extract given with CdCl<sub>2</sub> caused a lessening in MDA, increment in SOD in liver. Similar outcomes were gotten by the finding with (Rasoolijazi *et al.*, 2015 and Sakr *et al.*, 2015)

Liver damage was controlled by surveying serum levels of TNF- $\alpha$  and IL- $\beta$ . In our examination, hepatotoxicity actuated by AFB1 was reflected by a stamped height of TNF- $\alpha$  and IL- $\beta$  exercises. These outcomes are in concurrence with the aftereffects of numerous examinations many studies (Marin *et al.*, 2002; Serhan, 2007; Dönmez and Keskin, 2008 and Weaver *et al.*, 2013). The systemic inflammatory reaction is interceded by actuated pro-inflammatory cytokines, for example, tumor putrefaction factor- $\alpha$  (TNF- $\alpha$ ), interleukins (IL-1 $\beta$  and IL-6) and oxygen radicals which may sharpen hepatocytes to the toxicity (Li *et al.*, 2010b).

In the current study, TNF- $\alpha$  level was reduced significantly in animals receiving thyme with AFB1, the effect which probably related to the carvacrol that has anti-inflammatory effect, carvacol was a known component presented in the thyme extract (Juhas *et al.*, 2008). Fachini-Queiroz *et al.*, (2012) said that the inhibitory impact of carvacol on leukocyte movement adds to its calming activity, notwithstanding the aggravation impact of thymol. Other study by (Elhabazi *et al.*, 2003) showed that thyme extract increase the quantity of polymorphonuclears and total lymphocytes.

Soosani and Sazegar, (2018) reported that *Thyme daenensis* extracts have cell reinforcement and calming properties and can enhance liver damage in mice by diminishing pro-inflammatory TNF and IL-6 cytokines. Thus, this concentrate may be utilized as anti-inflammatory and hepato-protective agent.

Nafees *et al.*, (2016) reported that extract of Thyme leaves has been shown to reduce oxidative stress and liver inflammation induced by aflatoxine in Wistar rats. Abdel-Aziem *et al.*, (2014) indicated treatment with Thyme led to

significant decrease in TNF- $\alpha$  and IL-6 levels compared with the control group. Ahmed *et al.*, (2015) report that the aflatoxicated rats that treated with rosemary and thyme showed a significant diminution the levels of aflatoxin residues in testis, seminal vesicle and prostate gland.

## CONCLUSION

This study suggests the potent role of Thyme and rosemary in management of liver toxicity -induced by Aflatoxin and this effect could be attributed to their antioxidant activity.

## REFERENCES

- Abd El-Ghany, M.A.; Motawee, M.M.; El-Kewawy, H.E. (2012): Biological effects of yoghurt with rosemary on injured liver rats. *Aust. J. Basic Appl. Sci.* 6:525–532.
- Abd El Kader, M.A. and Mohamed, N.Z. (2012): Evaluation of Protective and Antioxidant Activity of Thyme (*Thymus Vulgaris*) Extract on Paracetamol-Induced Toxicity in Rats, *Australian Journal of Basic and Applied Sciences*, 6(7):467-474.
- Abdel-Aziem, S.H.; Hassan, A.M.; El-Denshary, E.S.; Hamzawy, M.A.; Mannaa, F.A. and Abdel-Wahhab, M.A. (2014): Ameliorative effects of thyme and calendula extracts alone or in combination against aflatoxins-induced oxidative stress and genotoxicity in rat liver. *Cytotechnology*; 66:457–70.
- Abdel-Wahhab, K.; El-Shamy, K.; El-Beih, N.; Morcy, F. and Mannaa, F. (2011): Protective effect of a natural herb (*Rosmarinus officinalis*) against hepatotoxicity in male albino rats. *Comunicata Scientiae*; 2(1):9-17.
- Abdel-Wahhab, M.A.; Ahmed, H.H. and Hagazi, M.M. (2006) : Prevention of aflatoxin B1-initiated hepatotoxicity in rat by marine algae extracts . *J Appl Toxicol* 26:229-38.
- Abnet, C.C. (2007): Carcinogenic food contaminants. *Cancer Invest*, 25: 189-196.
- Ahmed, N.H.; El-Mokhtar, N.M. and Abd El-Aal, R.A. (2015): Assessing the effect of Thyme and Rosemary as antiaflatoxicosis on fertility in male rats. *J Am Sci*; 11(12):294-302.
- Al-Attar, A.M. and Shawush, N.A. (2015): Influence of olive and rosemary leaves extracts on chemically induced liver cirrhosis in male rats. *Saudi J Biol Sci*; 22(2):157-63.
- Al-Kattan, M.M.; Abdul-Fattah, H.J. and Al-Annaz, R.M. ( 2012): Effect of Salvia Powder on Blood and Histological Picture of Liver and Heart Muscle in Male Albino Mice, *College of Science, University of Mosul, RSM*, 23(2): 1-14.
- Al-Malki , A.L. (2010): Antioxidant properties of thymol and butylated hydroxytoluene in carbon tetrachloride-induced mice liver injury. *JKAU Sci*; 22:239–48.

- Al-Sereiti, M.R.; Abu-Amer, K.M. and Sen, P. (1999): Pharmacology of Rosemary (*Rosmarinus officinalis* Linn.) and Its Therapeutic Potentials. *Indian Journal of Experimental Biology*, 37:124-130.
- Amarowicz, R.; Zegarska, Z.; Rafałowski, R.; Pegg, R.B.; Karamac, M. and Kosin, A. (2009): Antioxidant activity and free radical-scavenging capacity of ethanolic extracts of thyme, oregano, and marjoram, *Eur. J. Lipid Sci. Technol.*, 110(1):1-7.
- Andallu, B. and Vardacharyulu, N. (2001): Effect of mulberry leaves on diabetes. *Int. J. Diab. Dev. Countries*, 21: 147-151.
- Aristatile, B.; Al-Numair, K.S.; Veeramani, C. and Pugalendi, K.V. (2009): Effect of carvacrol on hepatic marker enzymes and antioxidant status in D-galactosamine-induced hepatotoxicity in rats. *Fundam Clin Pharmacol*;23: 757-65.
- Awad, M.H.; Atta, A.; Abdel Ghany, M.; and et al., (2011): Effect of a Specific Combination of Mannan-Oligosaccharides and  $\beta$ -Glucans Extracted from Yeast Cells Wall on the Health Status and Growth Performance of Ochratoxicated Broiler Chickens. *J. of American Science*, 7 (3):82-96.
- Beena, K.D. and Rawat, D.S. (2013): Synthesis and antioxidant activity of thymol and carvacrol based Schiff bases. *Bioorg Med Chem Lett.* 1;23(3):641-5.
- Beutler, E.; Dubon, B. and Kelly, M. (1963): Improved method for determination of blood Catalase. *J. Lab. Clin. Med.*, 61: 882-888.
- Bozin, B.; Mimica-Dukic, N.; Simin, N. and Anackov, G. (2006): Characterization of the volatile composition of essential oils of some lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils, *J Agric Food Chem*, 54(5):1822-1828.
- Del Bano, M.J.; Castillo, J.; Garcia, O.B.; Lorente, J.; Martin-Gil, R.; Acevado, C. and Alcaraz, M. (2006): Radioprotective-antimutagenic effects of rosemary phenolics against chromosomal damage induced in human lymphocytes by gamma-rays. *J Agric. Food Chem*, 54(6): 2064-2068.
- Domaracky, M.; Rehak, P.; Juhas, S. and Koppel, J. (2007): Effects of selected plant essential oils on the growth and development of mouse pre-implantation embryos *in vivo*, *Physiol. Res.*, 56(1):97-104.
- Dönmez, N. and Keskin, E. (2008): The effects of aflatoxin and glucomannan on some antioxidants and biochemical parameters in rabbits. *Acta Vet. Beograd*, 58: 307-313.
- Draper, H. H. and Hadley, M. (1990): Malondialdehyde determination as index of lipid peroxidation. *Methods Enzymol*, 186: 421-431.
- Eaton, D.L. and Gallagher, E.P. (1994): Mechanisms of aflatoxin carcinogenesis. *Annu Rev Pharmacol Toxicol*; 34:135-72.
- Elhabazi, K.; Dicko, A.; Desor, F.; Dalal, A.; Younos, C. and Soulimani, R. (2003): Preliminary study on immunological and behavioural effects of *Thymus broussonetii* Boiss, an endemic species in Morocco, *Mediators Inflamm.*, 12(6):323-328.
- El-Nekeety, A.A.; Mohamed, S.R.; Hathout, A.S.; Hassan, N.S.; Aly, S.E. and Abdel-Wahhab, M.A. (2011): Antioxidant properties of *Thymus vulgaris* oil against aflatoxin-induced oxidative stress in male rats, *Toxicol*, 57(7):984-991.
- Fachini-Queiroz, F.C.; Kummer, R.; Estevão-Silva, C.F.; Carvalho, M.; Cunha, J.M.; Grespan, R.; Bersani-Amado, C.A. and Cuman, R. (2012): Effects of Thymol and Carvacrol, Constituents of *Thymus vulgaris* L. Essential Oil, on the Inflammatory Response, Evidence-based Complementary and Alternative Medicine, 657026.
- Gong, Y.; Hounsa, A.; Egal, S.; Turner, P.C.; Sutcliffe, A.E.; Hall, A.J.; Cardwell, K. and Wild, C.P. (2004): Post-weaning exposure to aflatoxin results in impaired child growth: a longitudinal study in Benin. *Environ. Health Perspect*, 112: 1334-1338.
- Guarisco, J.A.; Hall, J.O. and Coulombe, R.A. (2008): Mechanisms of butylated hydroxytoluene chemoprevention of aflatoxicosis-inhibition of aflatoxin B(1) metabolism. *Toxicol Appl Pharmacol* 227:339-46.
- Hamzawy, M.A.; El-Denshary, E.S.; Hassan, N.S.; Manaa, F. and Abdel-Wahhab, M.A. (2012): Antioxidant and hepatoprotective effects of *Thymus vulgaris* extract in rats during aflatoxicosis. *Global J Pharmacol*;6(2):106-17.
- Haroun, E.M.; Mahmoud, O.M. and Adam, S.E. (2002): Effect of feeding cuminum cyminum fruits, *Thymus Vulgaris* leaves or their or mixture to rats, *Vet. Hum. Toxicol*, 44(2):67-9.
- Höferl, M.; Krist, S. and Buchbauer, G. (2006): Chirality influences the effects of linalool on physiological parameters of stress. *Planta Med.* 72(13):1188-1192.
- Juhas, S.; Bujnakova, D.; Rehak, P.; Čikos, Š; Czikkova, S; Vesela, J.; Ilkova, G. and Koppel, J. (2008): Anti-Inflammatory Effects of Thyme Essential Oil in Mice, *Acta Vet.*, 77:327-334.
- Kadri, A.; Zarai, Z.; Ben Chobba, I.; Bekir, A.; Gharsallah, N.; Damak, M. and Gdoura, R. (2011): Chemical constituents and antioxidant properties of *Rosmarinus officinalis* L. essential oil cultivated from South-Western Tunisia. *J. Med. Plants. Res.* 5:5999-6004.
- Kakkar, P.; Das, B. and Synder, P.N. (1984): A modified spectrophotometric assay of superoxide dismutase. *Ind. J. Biochem. Biophys*, 21: 131-132.
- Kamkar, A.; Yazdankhah, S.; Mohammadi Nafchi, A. and MozaffariNejad, A.S. (2013): Aflatoxin M1 in raw cow and buffalo milk in Shush city of Iran. *Food Add Contamin: Part B*.
- Kim, J.H.; Jeong, Y.J.; Hong, J.M.; Kim, H.R.; Kang, J.S.; Lee, W.J. and Hwang, Y.I. (2014): Chronic vitamin C insufficiency aggravated thioacetamide -induced liver fibrosis in gulo-knockout mice. *Free Radic. Biol. Med.* 67:81-90.
- Kodama, M.; Inoue, F. and Akao, M. (1990): Enzymatic and non -enzymatic formation of free radicals from aflatoxin B1. *Free Radic Res Commun*; 10:137-42.

- Li, G.; Jiang, W.; Tian, J.; Qu, G.; Zhu, H. and Fu, F. (2010a): In vitro and in vivo antifibrotic effects of rosmarinic acid on experimental liver fibrosis. *Phytomedicine* 17:282–288.
- Li, J.; Zhu, X.; Liu, F.; Cai, P.; Sanders, C. and Lee, W.M. (2010b): Cytokine and autoantibody patterns in acute liver failure. *J Immunotoxicol*; 7(3):157-64.
- Machado, D.G.; Cunha, M.P.; Neis, V.B.; Balen, G.O.; Colla, A.; Bettio, L.E.; Oliveira, A.; Pazini, F.L.; Dalmarco, J.B.; Simionatto, E.L.; Pizzolatti, M.G. and Rodrigues, A.L. (2013): Antidepressant-like effects of fractions, essential oil, carnosol and betulinic acid isolated from *Rosmarinus officinalis* L. *Food Chem.* 136, 999–1005.
- Mansour, H.A.; Newairy, A.A.; Yousef, M.I. and Sheweita, S.A. (2002): Biochemical study on the effects of some Egyptian herbs in alloxan-induced diabetic rats, *Toxicology*, 170 (3), 221-228.
- Marin, D.E.; Taranu, I.; Bunaciu, R.P.; Pascale, F.; Tudor, D.S.; Avram, N.; Sarca, M.; Cureu, I.; Criste, R.D.; Suta, V. and Oswald I.P. (2002): Changes in performance, blood parameters, humoral and cellular immune responses in weanling piglets exposed to low doses of aflatoxin. *J. Anim. Sci.*, 80: 1250-1257.
- Munné-Bosch, S. and Alegre, L. (2001): Subcellular compartmentation of the diterpene carnosic acid and its derivatives in the leaves of rosemary. *Plant physiology*, 125(2):1094-1102.
- Nabavi, S. F.; Tenore, G. C.; Daglia, M.; Tundis, R.; Loizzo, M. R. and Nabavi, S. M. (2015): The cellular protective effects of rosmarinic acid: from bench to bedside. *Curr. Neurovasc. Res.*, 12(1): 98-105.
- Nafees, S.; Ahmad, S.T.; Arjumand, W.; Rashid, S.; Ali, N. and Sultana, S. (2013): Carvacrol ameliorates thioacetamide-induced hepatotoxicity by abrogation of oxidative stress, inflammation, and apoptosis in liver of Wistar rats. *Hum Exp Toxicol*; 32(12):1292-304.
- Parmar, J.; Sharma, P.; Verma, P.; Sharma, P. and Goyal, P.K. (2011): Anti-tumor and anti-oxidative activity of *Rosmarinus officinalis* in 7,12 dimethyl benz (a) anthracene induced skin carcinogenesis in mice. *Am. J. Biomed. Sci.* 3:199–209.
- Peters, L.P. and Teel, R.W. (2003): Effect of high sucrose diet on liver enzyme content and activity and aflatoxin B1-induced mutagenesis. *In Vivo*, 17: 205-210.
- Rasoolijazi, H.; Mehdizadeh, M.; Soleimani, M.; Nikbakhte, F.; Eslami Farsani, M. and Ababzadeh, S. (2015): The effect of rosemary extract on spatial memory, learning and antioxidant enzymes activities in the hippocampus of middle-aged rats. *Med J Islam Repub Iran.* Vol. 29:187.
- Razavi-Azarkhiavi, K.; Behravan, J.; Mosaffa, F.; Sehatbakhsh, S.; Shirani, K. and Karimi, G. (2014): Protective effects of aqueous and ethanol extracts of rosemary on H2O2-induced oxidative DNA damage in human lymphocytes by comet assay. *J. Complement. Integr. Med.*, 11(1): 27-33.
- Reeves, R.G.; Nielsen, F.H. and Fahey, G.C. (1993): AIN-93 Purified Diets for Laboratory Rodents. *J. Nutr.* 123(1):1939-1951.
- Reddy, K.R.; Reddy, C.S. and Muralidharan, K. (2009a): Detection of *Aspergillus* spp. and aflatoxin B1 in rice in India. *Food Microbiology*. 26: 27-31.
- Reddy, K.R.N.; Reddy, C.S. and Muralidharan, K. (2009b): Potential of botanicals and biocontrol agents on growth and aflatoxin production by *Aspergillus flavus* infecting rice grains. *Food Control* 20: 173-178.
- Reitman, S. and Frankel, S.A. (1957): Colorimetric method for determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. *Am J Clin Path.* 28:56–63.
- Rasooli, I.; Rezaei, M.B. and Allameh, A. (2006): Ultra structural studies on antimicrobial efficacy of thyme essential oils on *Listeria monocytogenes*, *International Journal Infectious Diseases*, 10(4):236-241.
- Rubió, L.; Serra, A.; Chen, C.Y.; Macià, A.; Romero, M.P. and Covas, M.I. (2014): Effect of the co-occurring components from olive oil and thyme extracts on the antioxidant status and its bioavailability in an acute ingestion in rats. *Food Funct*; 5:740–7.
- Saito, Y.; Shiga, A.; Yoshida, Y.; Furuhashi, T.; Fujita, Y. and Niki, E. (2004): Effects of Novel Gaseous Antioxidative System Containing a Rosemary Extract on the Oxidation Induced by Nitrogen Dioxide and Ultraviolet Radiation. *Bioscience, Biotechnology, and Biochemistry*, 68:781-786.
- Sakr, S.A.; Bayomy, M.F. and El-Morsy, A.M. (2015): Rosemary extract ameliorates cadmium-induced histological changes and oxidative damage in the liver of albino rat. *The Journal of Basic & Applied Zoology*. 71:1–9.
- Salama, S.M.; Abdulla, M.A.; AlRashdi, A.S.; Ismail, S.; Alkiyumi, S.S. and Golbabapour, S. (2013): Hepatoprotective effect of ethanolic extract of *Curcuma longa* on thioacetamide induced liver cirrhosis in rats. *BMC Complement. Altern. Med.* 13:1–17.
- Samuel, S.M.; Aiko, V.; Panda, P. and Mehta, A. (2013): Aflatoxin B1 occurrence, biosynthesis and its degradation. *J Pure Appl Microbiol.*; 7: 965-971.
- Serhan, C.N. (2007): Resolution phase of inflammation: novel endogenous anti-inflammatory and pro-resolving lipid mediators and pathways, *Annual Review of Immunology*, 25:101–137.
- Seung, L.; Katumi, U.; Takayki, S. and Kwang-Geum, L. (2005): Identification of volatile components in basil and thyme leaves and their antioxidants properties, *Food. Chem.*, 91(1):131-137.
- Shanon, A.Q. (2011): The effect of *Zingiber officinale* and *Thyme Vulgaris* in the reproductive and productive performance of mothers broiler chickens (Ross 308), Faculty of Agriculture, University of Tikrit. (Ph.D thesis).
- Sharangi, A.B. and Guha, S. (2013): Wonders of leafy spices: Medicinal properties ensuring Human Health. *Science International*.; 312-317.

- Sharma, R. (2010): Recommendations on herbs and herbal formula in cancer prevention. Open Nutraceuticals J., 3: 129-140.
- Shen, HM. ; Shi, CY.; Shen, Y. and Ong, CN. (1996): Detection of elevated reactive oxygen species level in cultured rat hepatocytes treated with Aflatoxin B 1. Free Rad Biol Med; 21:139-46.
- Sherif, S.O.; Salama, E.E. and Abdel-Wahhab, M.A. (2009): Mycotoxins and child health: the need for health risk assessment. (Review) Int. J. Hyg. Environ. Health, 212: 347-368.
- Snedecor, G.W. and Cochran, W. (1980): Statistical methods., 7<sup>th</sup> Ed., Iowa State University Press, Ames, USA, Page 90.
- Soosani, B. and Sazegar, H. (2018): Effects of *Thymus daenensis* on inflammatory factors and liver toxicity induced by thioacetamide in rats. J Herbmed Pharmacol; 7(1):56-60.
- Tavakoli , H. R.; Kamkar , A . ; Riazipour, M.; Mozaffari Nejad, A .S, and Rafati , H. (2013) : Assessment of aflatoxin M1 levels by Enzyme-linked Immunosorbent Assay in yoghurt consumed in Tehran, Iran. Asian J Chem 25: 2836-2838.
- Umarani, M.; Shanthi, P. and Sachdanandam, P (2008): Protective effect of Kalpaamruthaa in combating the oxidative stress posed by aflatoxin B (1)-induced hepatocellular carcinoma with special reference to flavonoid structure-activity relationship. Liver Int 28:200-13.
- Virk, P.; Elobeid, M.; Hamad, S.; Korany, Z.; Al-Amin, M.; Daghestani, M.; Omer, S.; AlOlayan, E.; Siddiqui, M.I. and Mirghani, N.M. (2013): Ameliorative effects of *Emblica officinalis* and *Rosmarinus officinalis* on cadmium-induced oxidative stress in Wistar rats. J. Med. Plant Res. 7:805–818.
- Wathfeq, B.M.F.; Zaki, L.M.; Ibraheem, K.R. and Al-Rubaeem, S.H. (2011): Effect of *MenthapiperitaL.* Powder for different periods to the diet of broiler on some of biochemical characteristics, JAVS. 4(1).
- Weaver, A.C.; See, M.T.; Hansen, J.A.; Kim, Y.B; De Souza, A.L.P.; Teena, F.M. and Kim, S.W.( 2013): The use of feed additives to reduce the effects of aflatoxin and deoxynivalenol on pig growth, organ health and immune status during chronic exposure. Toxins, 5: 1261-1281.
- Weissman, N.; Schoenbach, E.B. and Armestead, E.B. (1950): The determination of sulfhydryl groups in serum. I. Methods and results on normal sera. J. biol. chem., 187:153.
- Williams, J.H.; Phillips, T.D.; Jolly, P.E.; Stiles, J.K.; Jolly, C.M. and Aggarwal, D. (2004): Human aflatoxicosis in developing countries; a review of toxicology, exposure, potential health consequences, and interventions. Am. J. Clin. Nutr., 80: 1106-1122.
- Wójtowicz-chomicz, K.; Stadnik, A.; Kowal, M.; Sztanke, K.; Sztanke, M. and borzęcki, A. (2011): disturbances of anti-oxidative balance in rats caused by aflatoxin b1. Bull Vet Inst Pulawy., 55:145-148.
- Zargar, S. (2014): Protective effect of *Trigonella foenum-graecum* on thioacetamide induced hepatotoxicity in rats. Saudi J. Biol. Sci. 21:139–145.

### تأثير الزعتر والروزماري على وظائف الكبد في الفئران المصابة بالتسمم بواسطة الأفلاتوكسين نيفين سيوفي اسماعيل قسم التغذية وعلوم الاطعمة- كلية الاقتصاد المنزلي- جامعة حلوان.

يلعب الزعتر والروزماري دوراً في حماية الكبد ، ويعملان كمضاد للتسمم. أجريت هذه الدراسة للتحقق من تأثير جرعة من الأفلاتوكسين في فئران التجارب والتنبؤ بأي فوائد أو أضرار من استخدام الزعتر والروزماري. تم تقسيم عدد 30 من ذكور الفئران البالغة من سلالة ألبينو إلى خمس مجموعات (كل مجموعة 6 فئران). تم تقسيم الفئران إلى مجموعتين رئيسيتين ، المجموعة الأولى الرئيسية (المجموعة الضابطة السالبة) ، تم تغذيتها على النظام الغذائي الأساسي فقط. المجموعة الرئيسية الثانية تم حقنها بالأفلاتوكسين ثم قسمت إلى أربع مجموعات فرعية على النحو التالي ، المجموعة الفرعية الأولى استخدمت كمجموعة ضابطة موجبة ، وتم تغذية المجموعات (3،2) على النظام الغذائي الأساسي المدعم مع الزعتر والروزماري عند مستوى 5 ٪ على التوالي. والمجموعة الرابعة تم تغذيتها على النظام الغذائي الأساسي والمدعم من خليط الزعتر والروزماري عند مستوى 2.5 ٪ لكل منهما. أشارت النتائج إلى أن التدعيم بالزعتر والروزماري وخليطهما أدت إلى تحسن كبير في وظائف الكبد وكذلك حدوث ارتفاع ذو دلالة احصائية  $p > 0.05$  في قيم الالبيومين ، الجلوبيولين والبروتين الكلي ، النشاط المضاد للأكسدة SOD و CAT وتنقص MDA بشكل معنوي . في حين أن الزعتر ، الروزماري وخليطها أدت الي حدوث انخفاض معنوي ( $P < 0.05$ ) في مستوى  $TNF-\alpha$  و  $IL-1-\beta$  مقارنة بالمجموعة الضابطة الموجبة. أوصت الدراسة الحالية باستخدام الزعتر والروزماري لحماية الجسم من التسمم الناتج عن الأفلاتوكسين .